

# Task 4: Novel bio-electro-mechanical tools

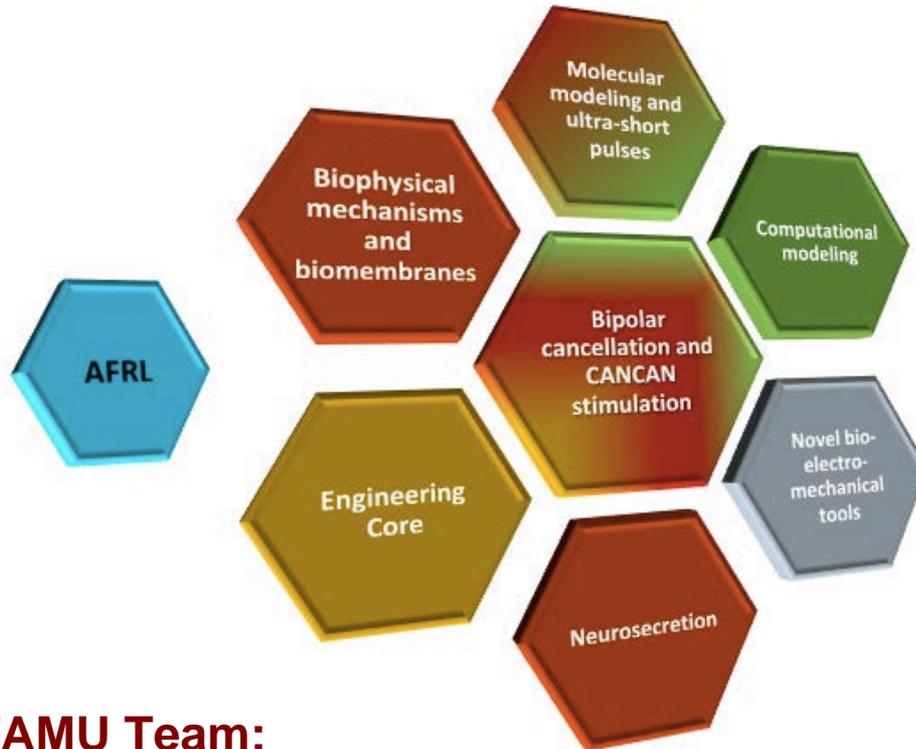
Nascent biophysical tools to elucidate nanoelectropulse-induced electromechanical interactions

Vladislav Yakovlev



TEXAS A&M  
UNIVERSITY.

# Acknowledgement



## **TAMU Team:**

Georgi Petrov, Charles Ballmann, Jon Thompson, Zhaokai Meng, Maria Troyanova-Wood, Zach Coker, Josh Lalonde, Mark Keppler, Chris and Kassie Marble, John Hernandez, Carlos Tovar, Grady Burnett, Nathan Arnold, Dr. Phil Hemmer

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Dr. Warwick Bowen



THE UNIVERSITY  
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AUSTRALIA



# Motivation

Electric field measurements across membrane

Local chemistry assessment

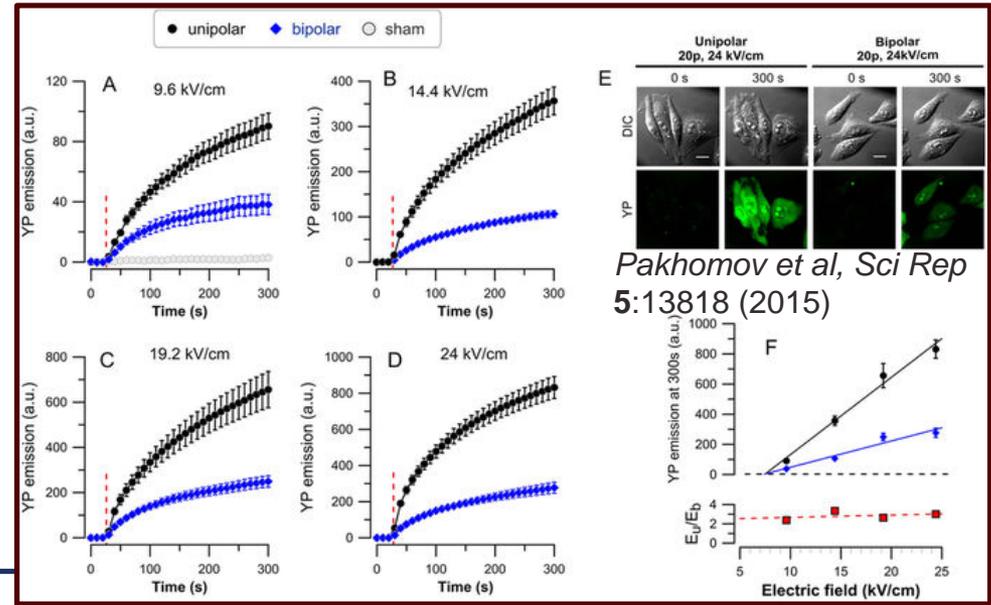
Local mechanical properties assessment

Mechanism?

- **assisted membrane discharge (?)** - need more v
- calcium in and out
- low frequency content
- **a 2-step process:**

- *oxidation (or some chemistry), then long-lasting permeabilization* - need experiments

- *"foot-in-the-door" occlusion of transient pores* - need experiments (GUV?)



Pakhomov et al, *Sci Rep* 5:13818 (2015)

ments and models!

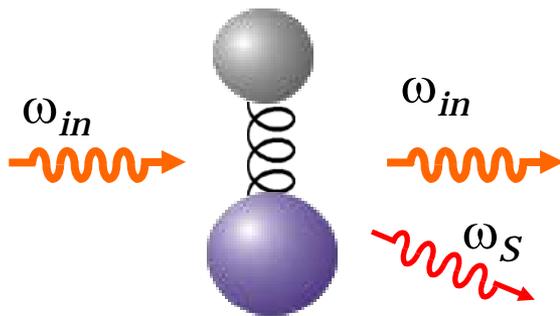
RS Son, KC Smith, TR Gowrishankar, PT Vernier, and JC Weaver. *J. Membr. Biol.* 247:1209-1228 (2014).



# Motivation

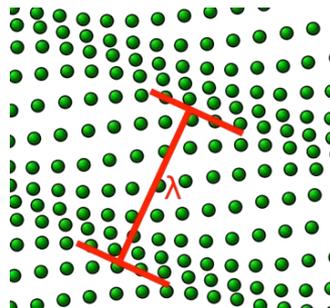
Develop **advanced** optical tools for **non-invasive** assessment of **electrical potentials** and mechanical properties (**elasticity and viscosity**).

## Raman spectroscopy



Molecular vibrations carry information about **structure and chemical composition**

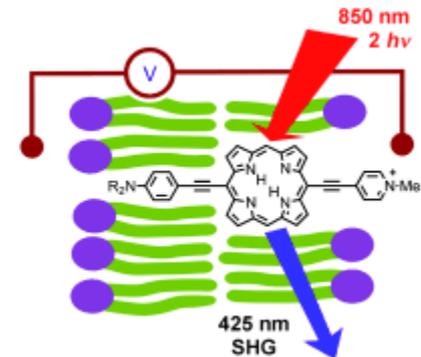
## Brillouin spectroscopy



$$\Delta\nu \propto \sqrt{K}$$

Acoustic oscillations carry information about **longitudinal modulus and viscosity**

## Other spectroscopies



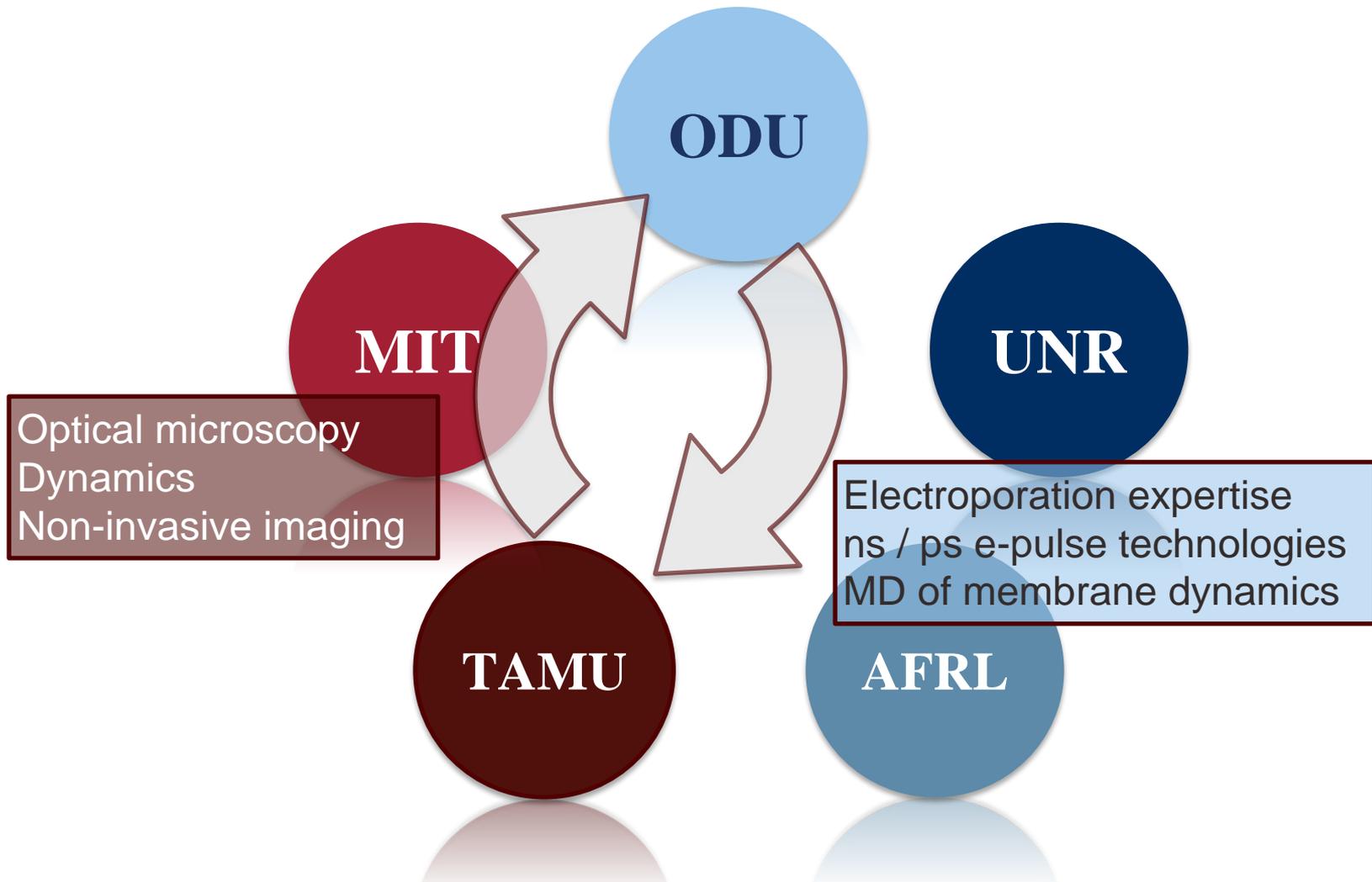
## True electro-optic effect(s)

$$\vec{E}(2\omega) = \chi^{(3)}(2\omega; \omega, \omega, 0) \vec{E}(\omega) \vec{E}(\omega) \vec{E}(0)$$

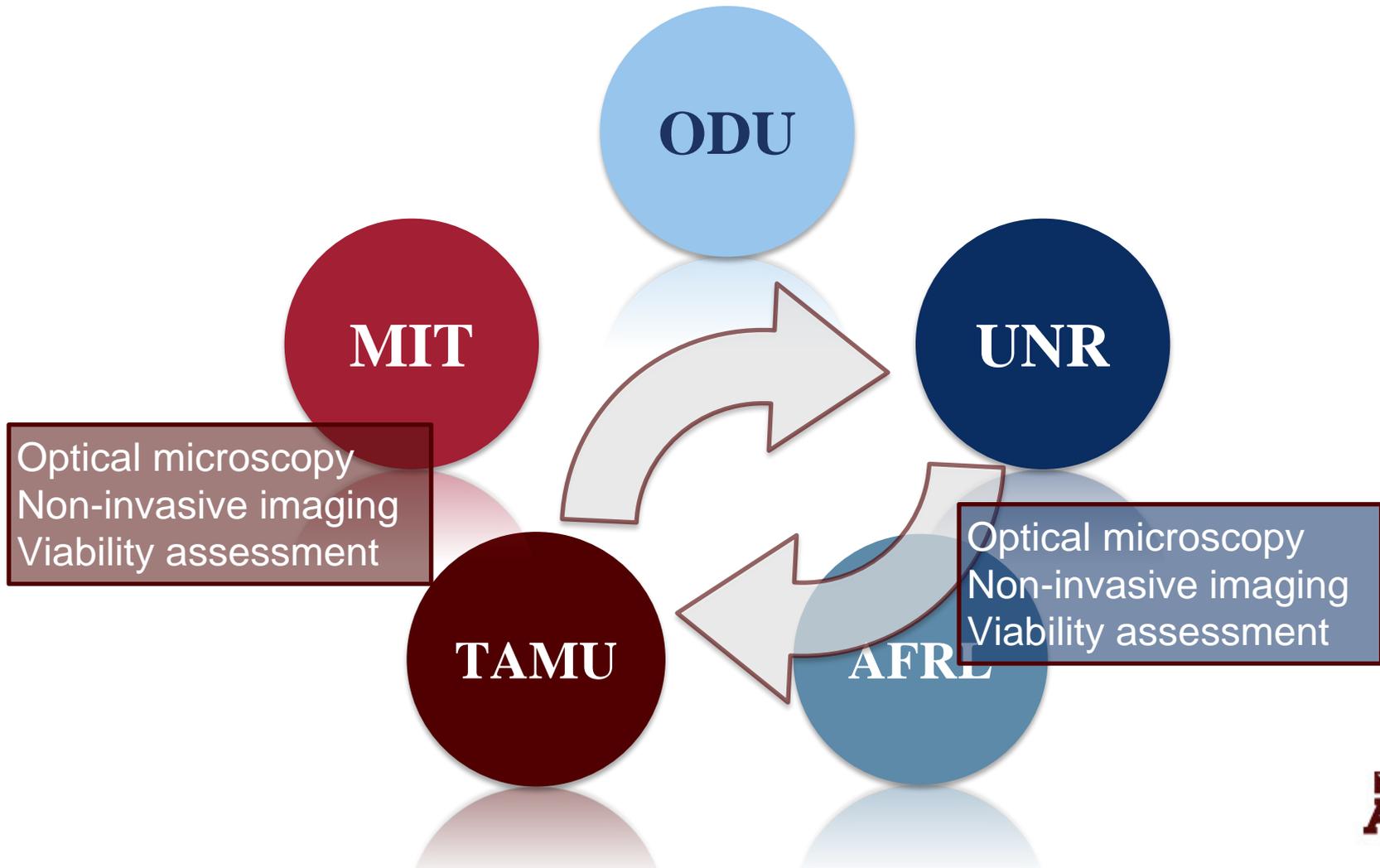
# Long-term goals

- Gain fundamental understanding of structure / property relationship on a nano/micro-scale and dynamical processes involved in nsPEF interaction with lipid membranes
- Develop new biophysical tools and instruments, which push sensitivity, molecular and structural specificity and spatial/temporal resolution
- Educate and train students and research personnel in a broad area of biophysics and bioengineering for DoD mission

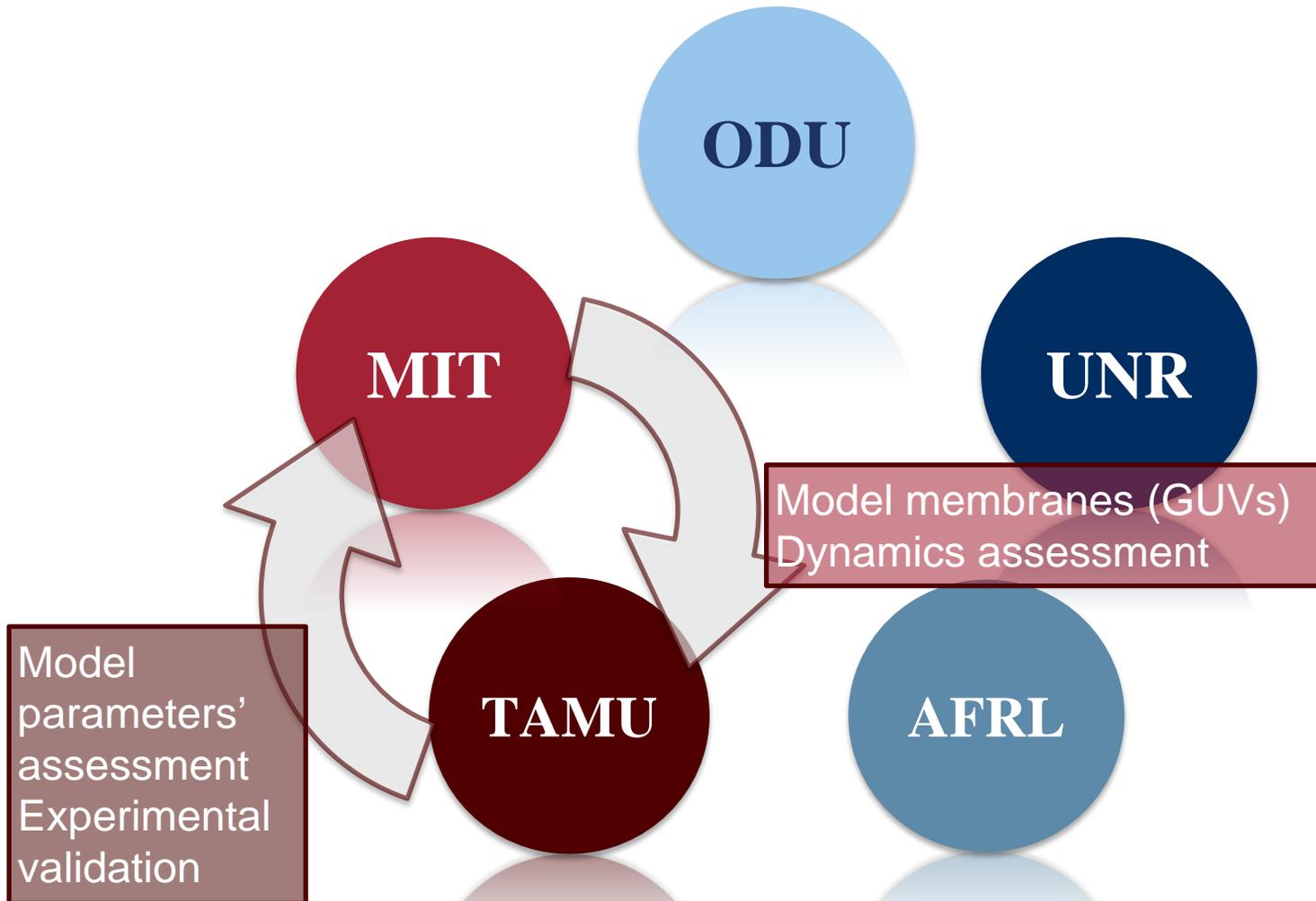
# Relationship to other tasks



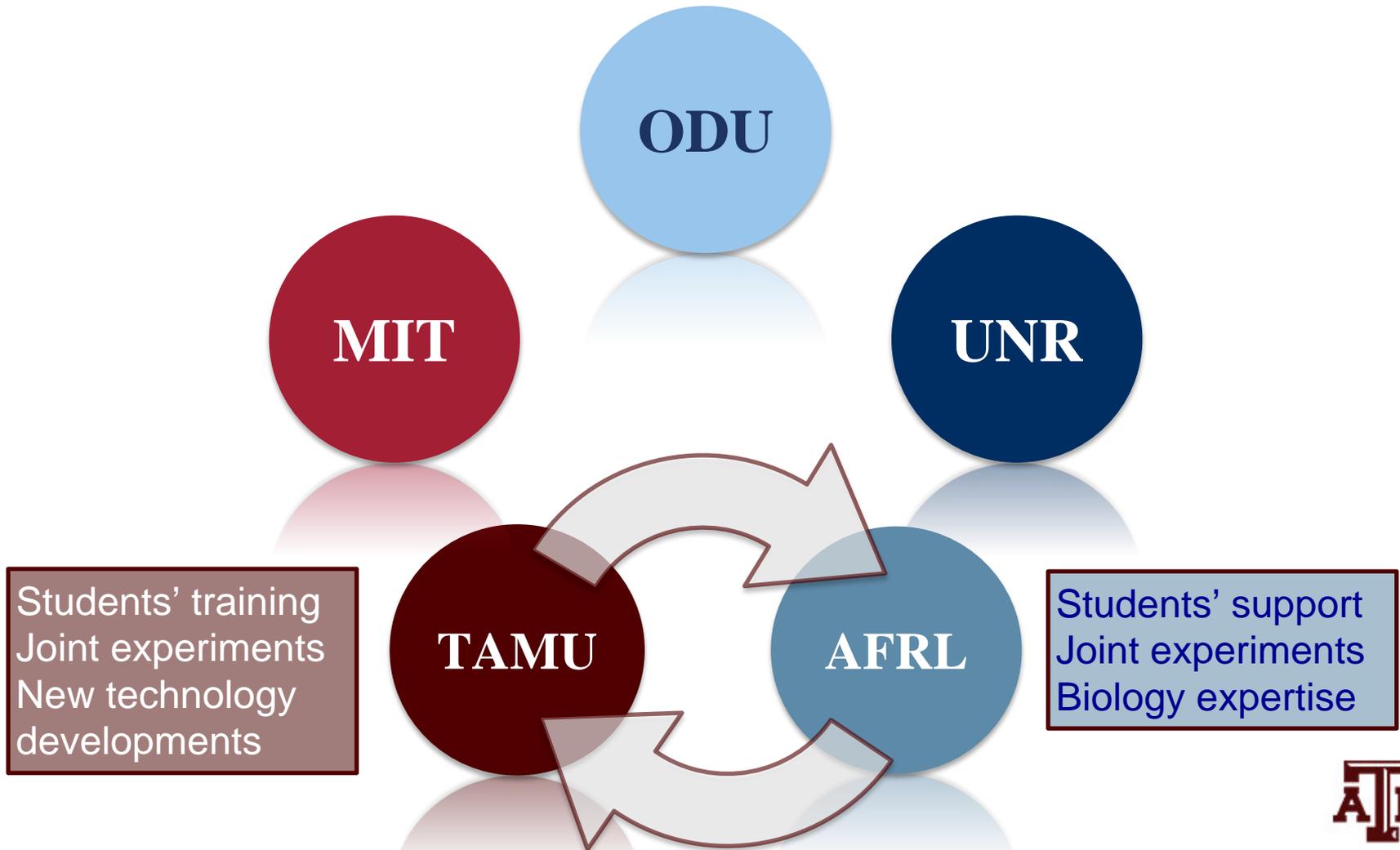
# Relationship to other tasks



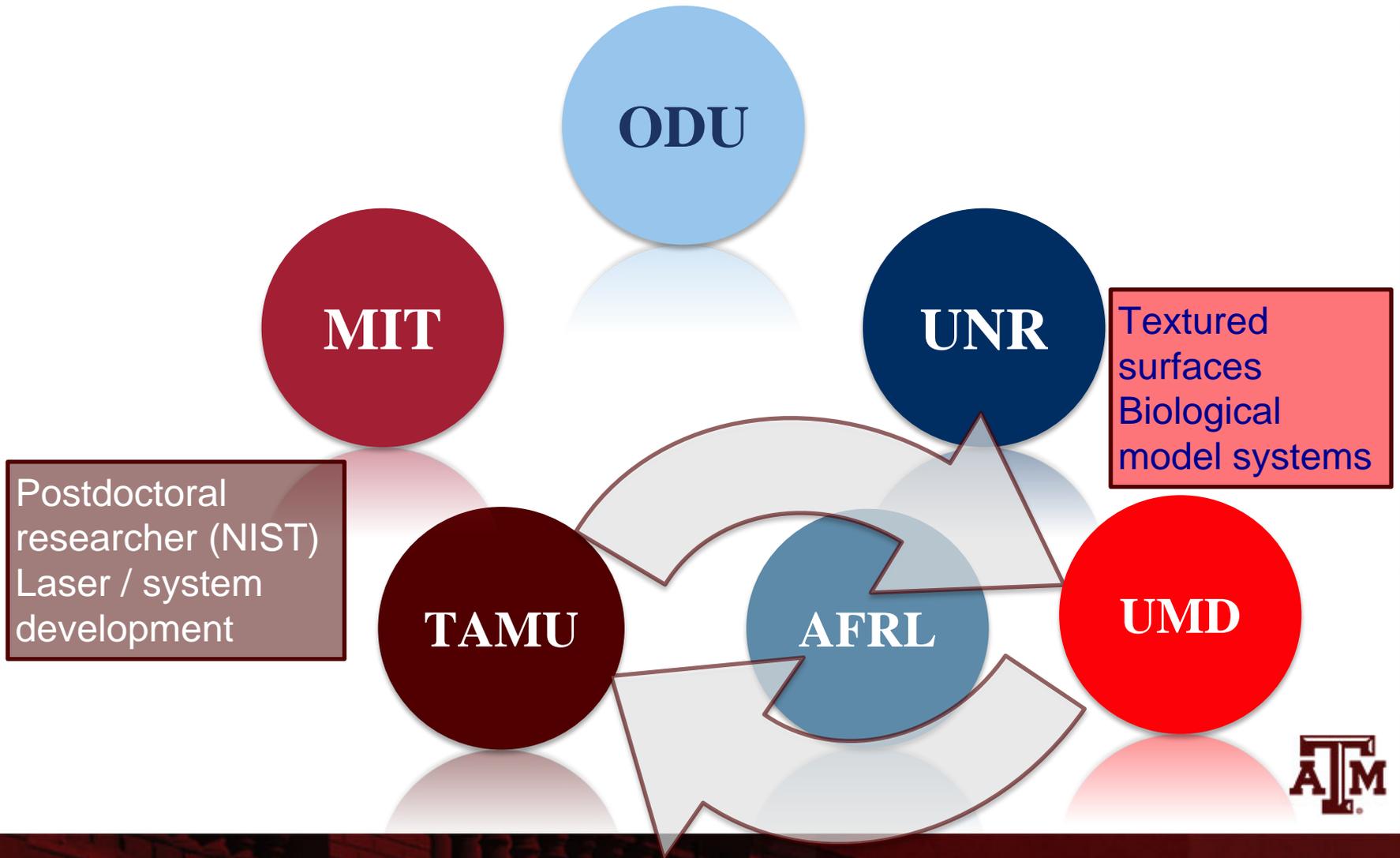
# Relationship to other tasks



# Relationship to other tasks



# Relationship to other MURI



# Impact

## Major scientific impact:

- (1) Developed instrument which allows microscopic mechanical assessment of living cells and tissues
- (2) Developed instrument which allows non-invasive measurements of electric potential across the membrane
- (3) Developed instrument which allows 1000x more accurate instrument to assess local chemical / structural composition

25 journal publications (+2 under review)

65 conference proceedings

34 conference presentations. including 1 Keynote, 1 Plenary, and 6 Invited

3 PhD students have graduated and are working for DoD

7 graduate students were involved in different aspects of the work

10 undergraduate students were involved

8 students worked as interns at AFRL



# Advancements and Challenges

## Successes and advancements

State of the art instrument for broadband coherent Raman imaging: unprecedented sensitivity to structure / chemical changes

State of the art instrument Brillouin imaging: unprecedented sensitivity to mechanical properties

Achieved the desired sensitivity of electric field imaging

Strong collaborative effort with the Air Force Research Laboratory

## Challenges and setbacks

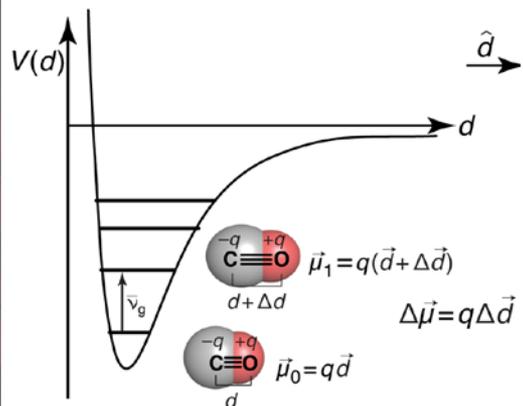
Imaging electrical fields potentials was found to be more challenging than it was thought at the time of proposal submission

Unexpected laser malfunction (now fixed) forced to explore some alternative strategies and techniques

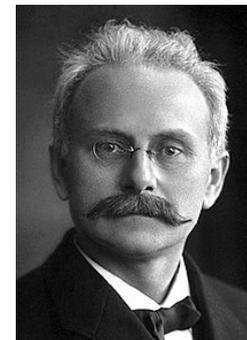
Initial hypothesis of signal significant enhancement via overtone resonant excitation only partially worked out: 20x signal enhancement, but the line-shapes were significantly distorted



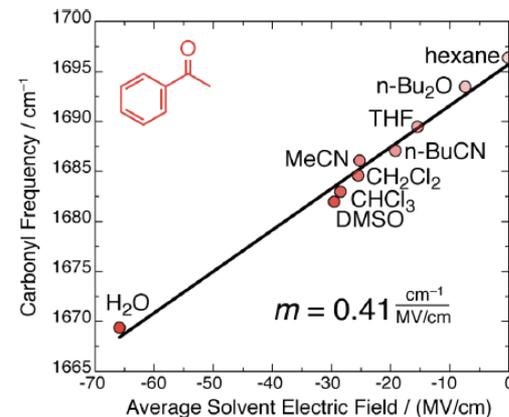
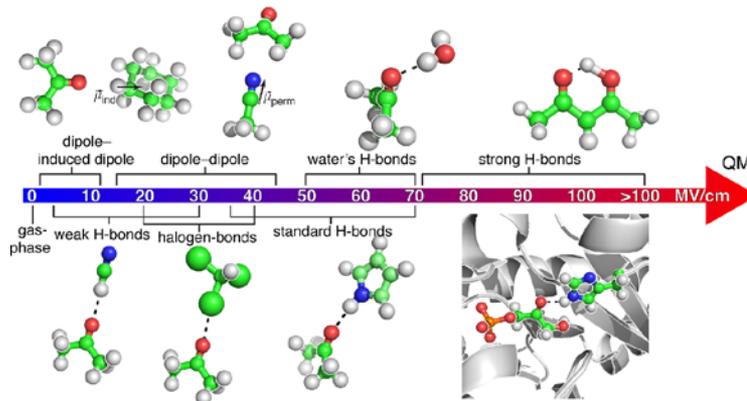
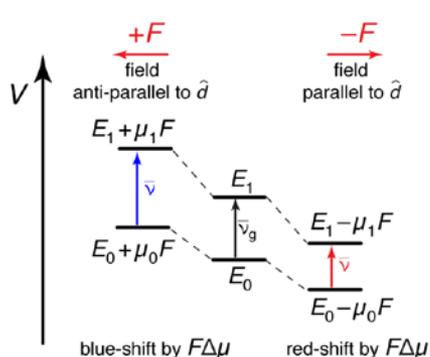
# Electrical potential measurements via vibrational Stark effect



Johanes Stark (Nobel Prize 1913)  
 Shifting and splitting emission lines in the presence of external electrical field. Considered one of the first verifications of quantum mechanics.



Vibrational Stark effect (quantum effect):  
**1 cm<sup>-1</sup>** shift per **MV/cm** electrical field

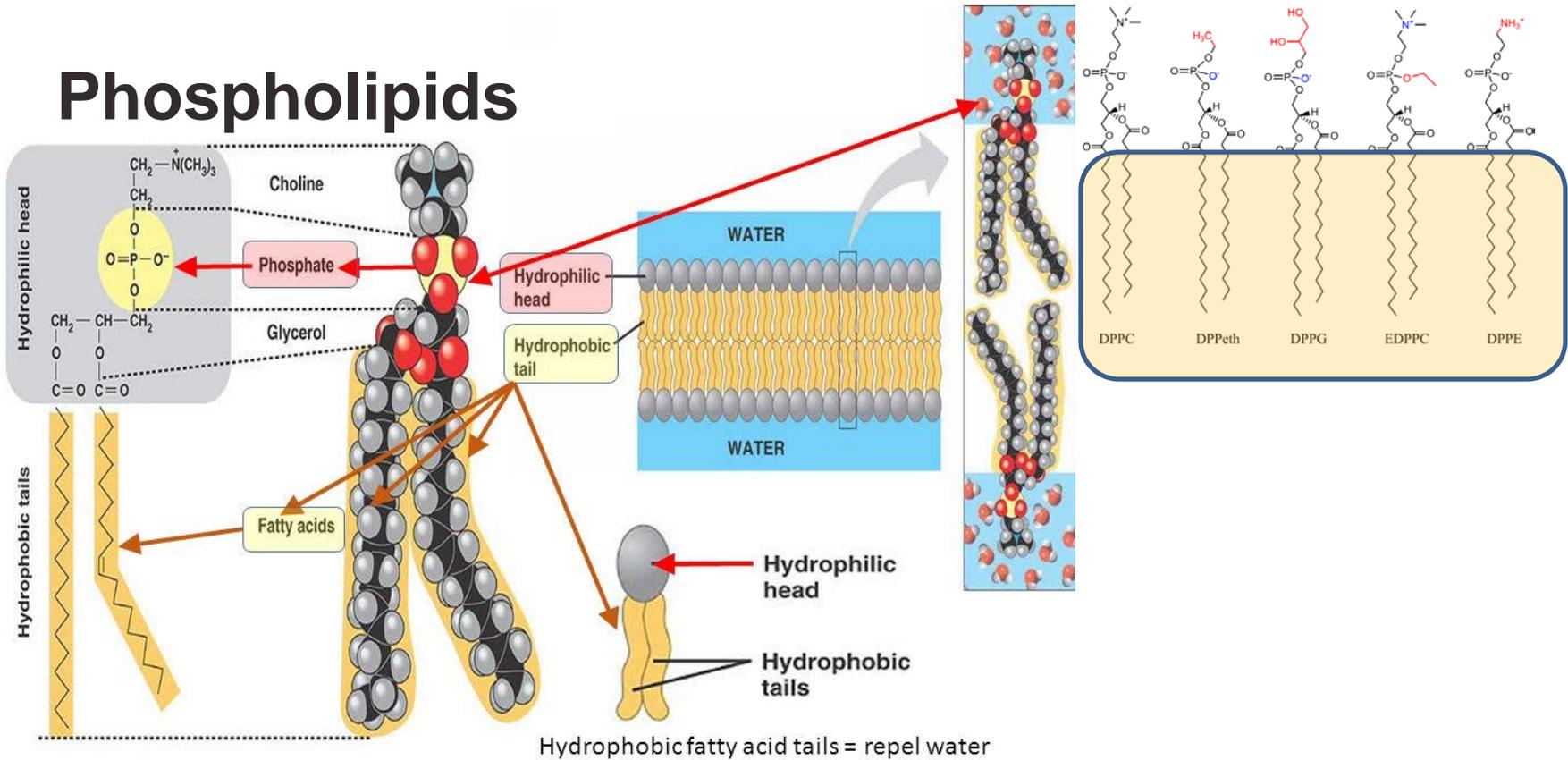


S. D. Fried, S. G. Boxer, "Measuring electric fields and noncovalent interactions using the vibrational stark effect," *Acc. Chem. Res.* 48(4): 998-1006 (2015).



# Structure of biological membrane

## Phospholipids



# Model system: giant unilamellar vesicle (GUV)

Biophysical Journal Volume 88 February 2005 1143-1155

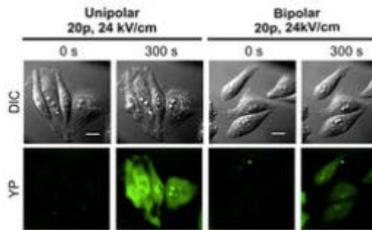
1143

## Electro-Deformation and Poration of Giant Vesicles Viewed with High Temporal Resolution

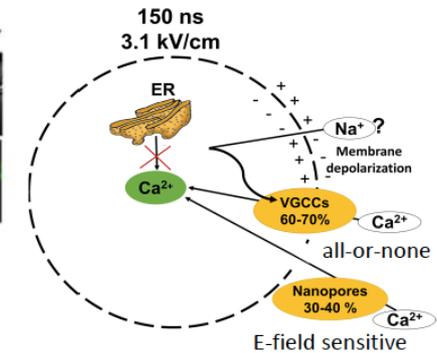
Karin A. Riske and Rumiana Dimova  
Max Planck Institute of Colloids and Interfaces, 14476 Golm, Germany

GUV provides a common motive for cellular membrane; it is simpler to model and is easier for interpretation, there are less unknown parameters; controllable shape, size and environment. It is also provides stable model for instrumentation validation.

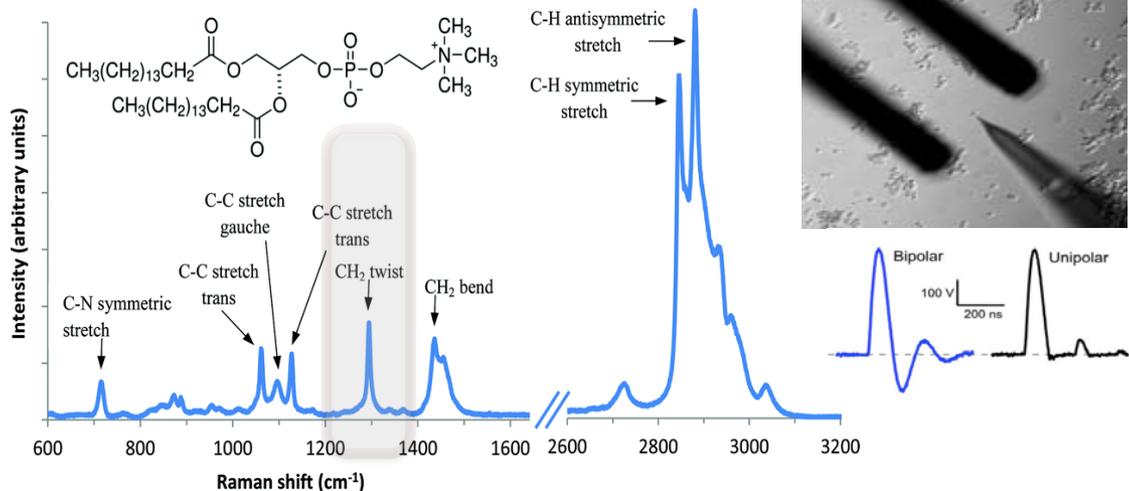
Pakhomov et al



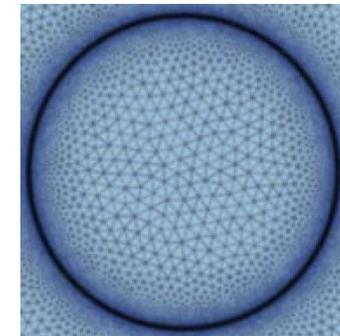
Craviso et al



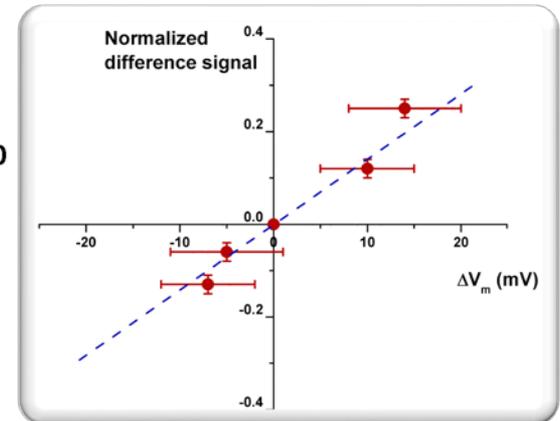
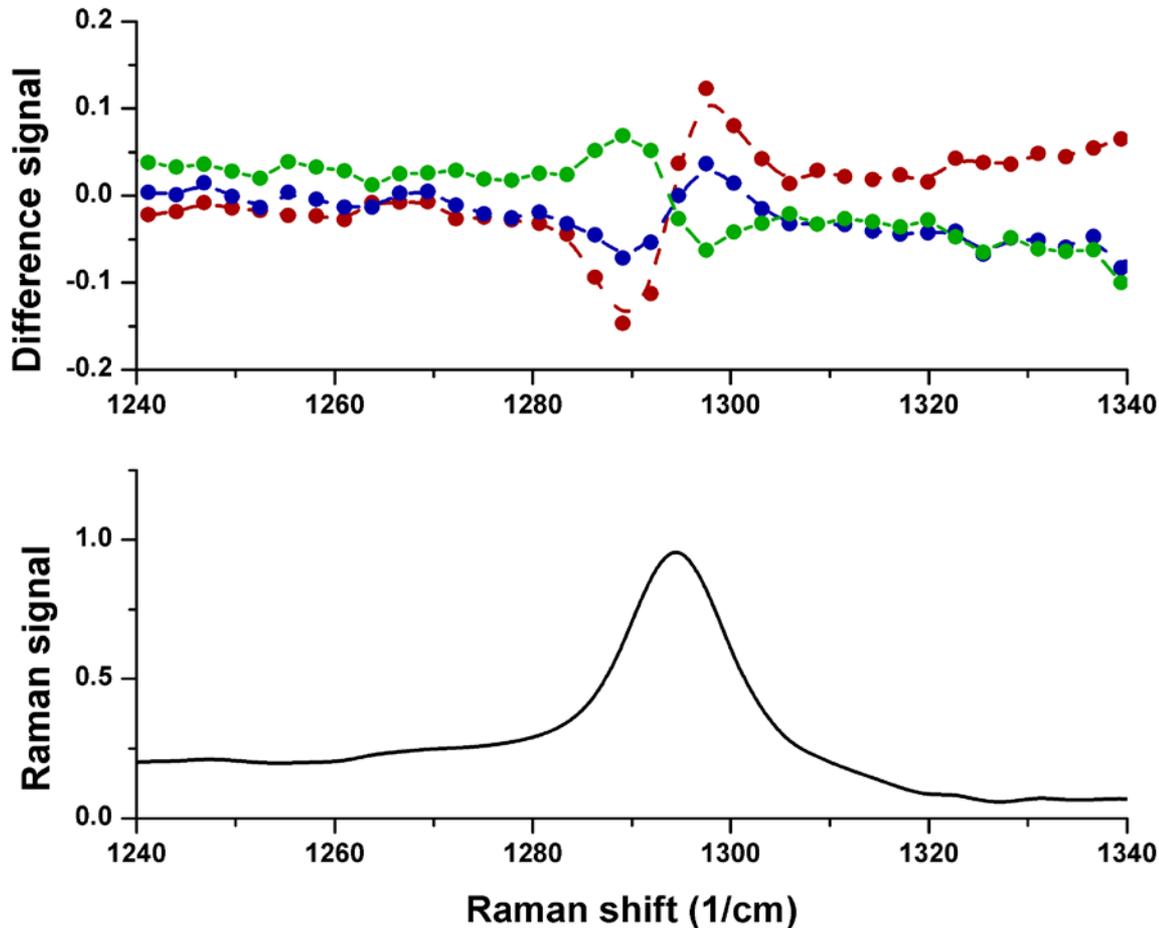
Technical core



Theory group



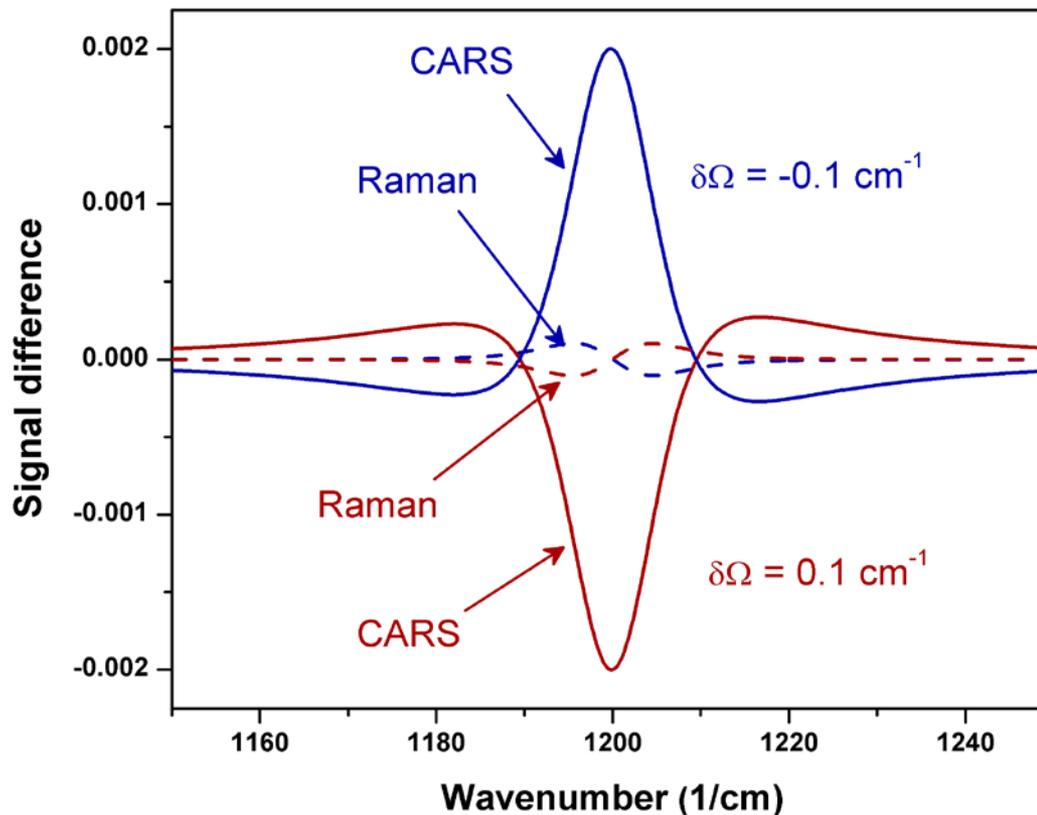
# Raman spectroscopy of GUV's



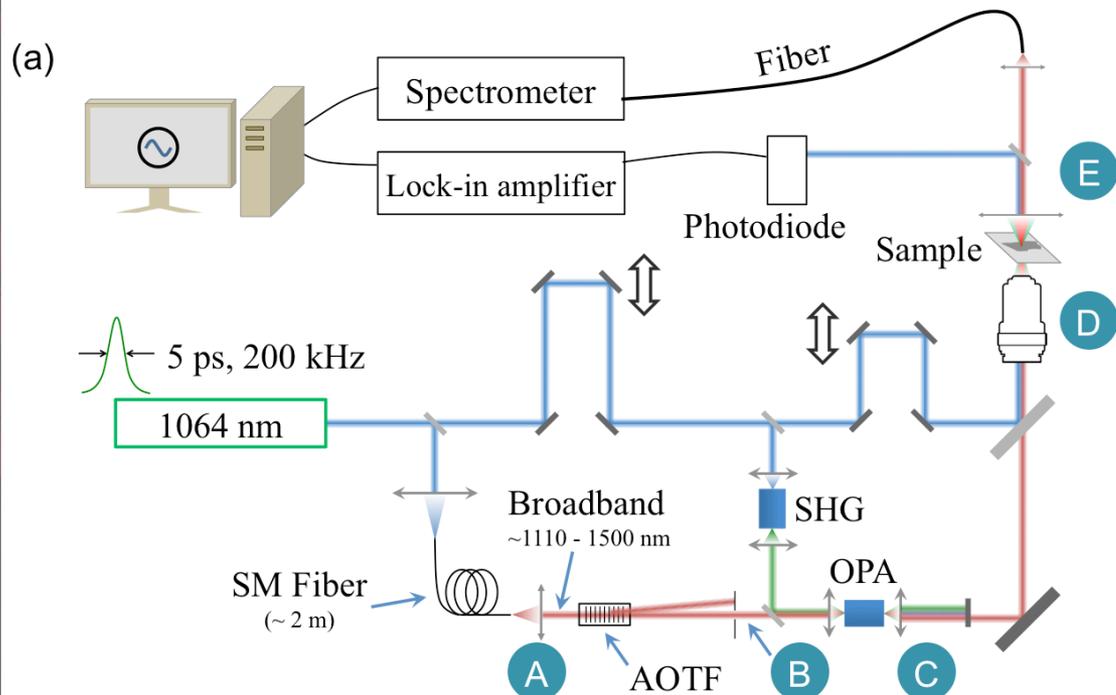
532-nm excitation, ~10 mW, 30-s acquisition time

# Raman vs CARS: Stark spectroscopy

Nor only CARS spectroscopy produces **stronger signal**, but it is also **stronger affected** by the frequency shift of vibrational transition



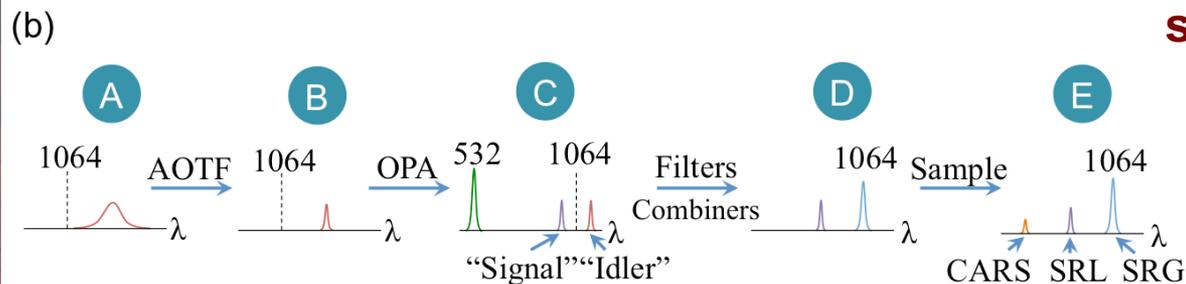
# Improved CARS/ SRS spectrometer



**No moving parts:**

wavelength selection is done through **continuously tunable AOTF**, and light is **amplified using OPA** to provide sufficient intensity and stability at the sample.

The wavelengths of both pump and Stokes beams are **derived from highly stabilized laser sources**



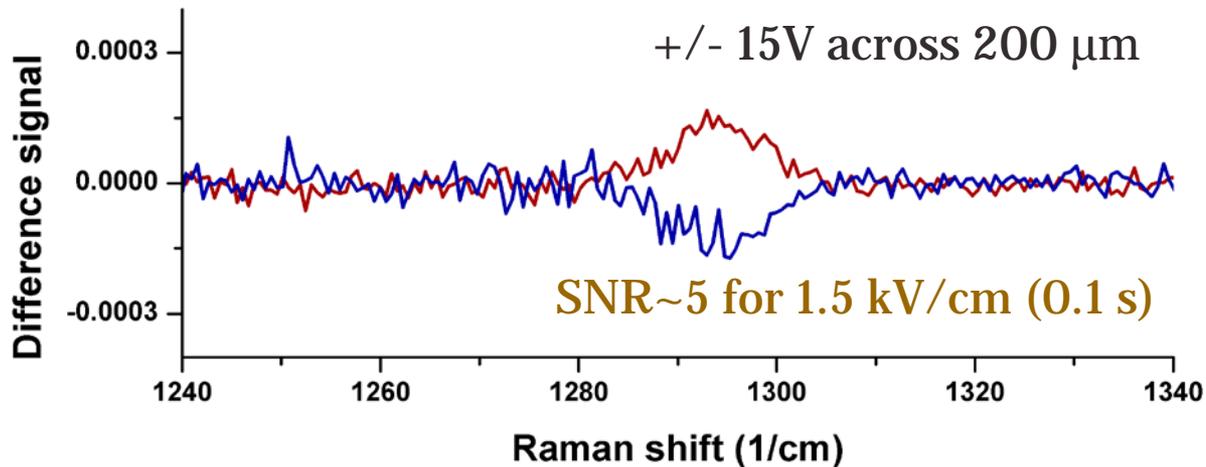
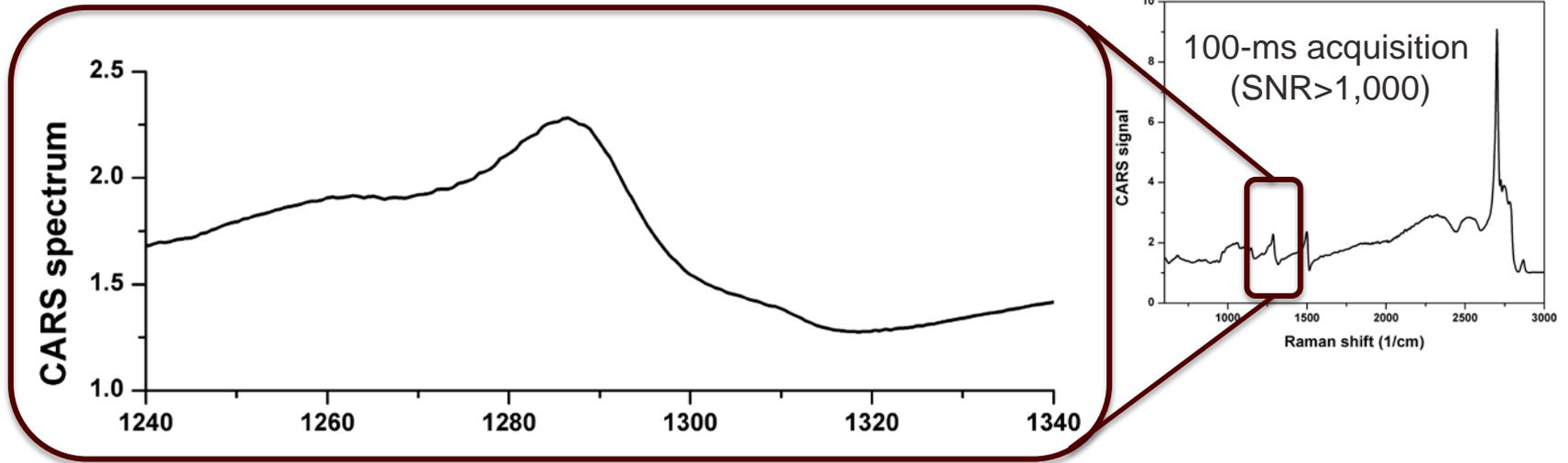
CW Ballmann, GI Petrov, VV Yakovlev, *Opt Lett* 42(1), 89-92 (2017); AJ Traverso et al *Light Sci Appl* 6(5), e16262 (2017).

Z Meng, GI Petrov, VV Yakovlev, *Sci Rep* 6, 20017 (2016)

GI Petrov, Z Meng, VV Yakovlev, *Opt Express* 23(19), 24669-24674 (2015)



# CARS Stark spectroscopy



# Mechanical assessment

Vol. 90, No. 2, 1979  
September 27, 1979

BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS  
Pages 656-662

Electrical potential affects rigidity (stiffness) of membranes

## POTENTIAL DEPENDENT RIGIDITY CHANGES IN LIPID MEMBRANE VESICLES

Peter I. Lelkes

Department of Membrane Research, The Weizmann Institute of Science,  
Rehovot, Israel

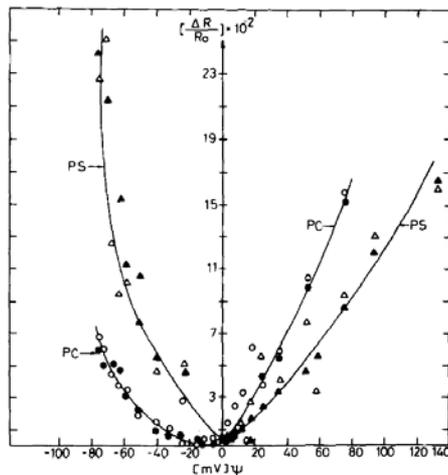
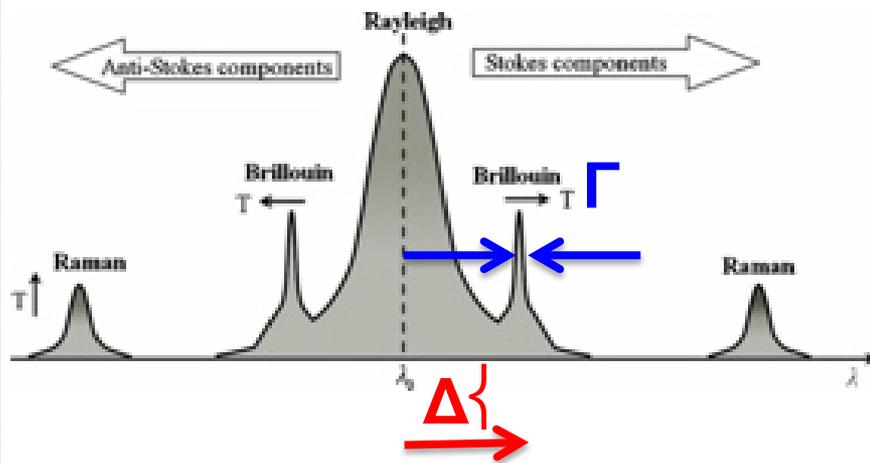
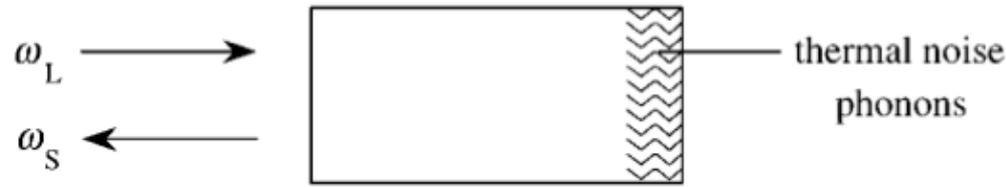


Figure 1: The relative change in membrane rigidity  $\Delta R/R_0$  in dependence of the transmembrane potential  $\psi$ , measured in phosphatidylcholine (PC) and phosphatidylserine (PS) vesicles. The different symbols represent independent experiments. For details, see text.

In this communication we present experimental evidence for a straightforward relationship between the transmembrane electrical potential and the membrane structure, expressed here as its rigidity. The biological implications of our results are significant and of general interest. A number of events on the cellular level are known to be initiated and/or accompanied by changes in the membrane potential, like ion-transport, hormonal action, enzyme-regulation, cell differentiation and proliferation, as well as cell growth and in some cases independently concomitant modifications in the membrane structure have been observed (25,26). Vertical translocations of proteins, and availability of receptor sites in biological membranes, which can be modulated by changes in the membrane fluidity (27), as well as by electrical fields across the membrane (28), underline the interrelation between membrane structure and transmembrane potential and its general occurrence in biological systems.

# Brillouin microscopy: viscoelastic assessment



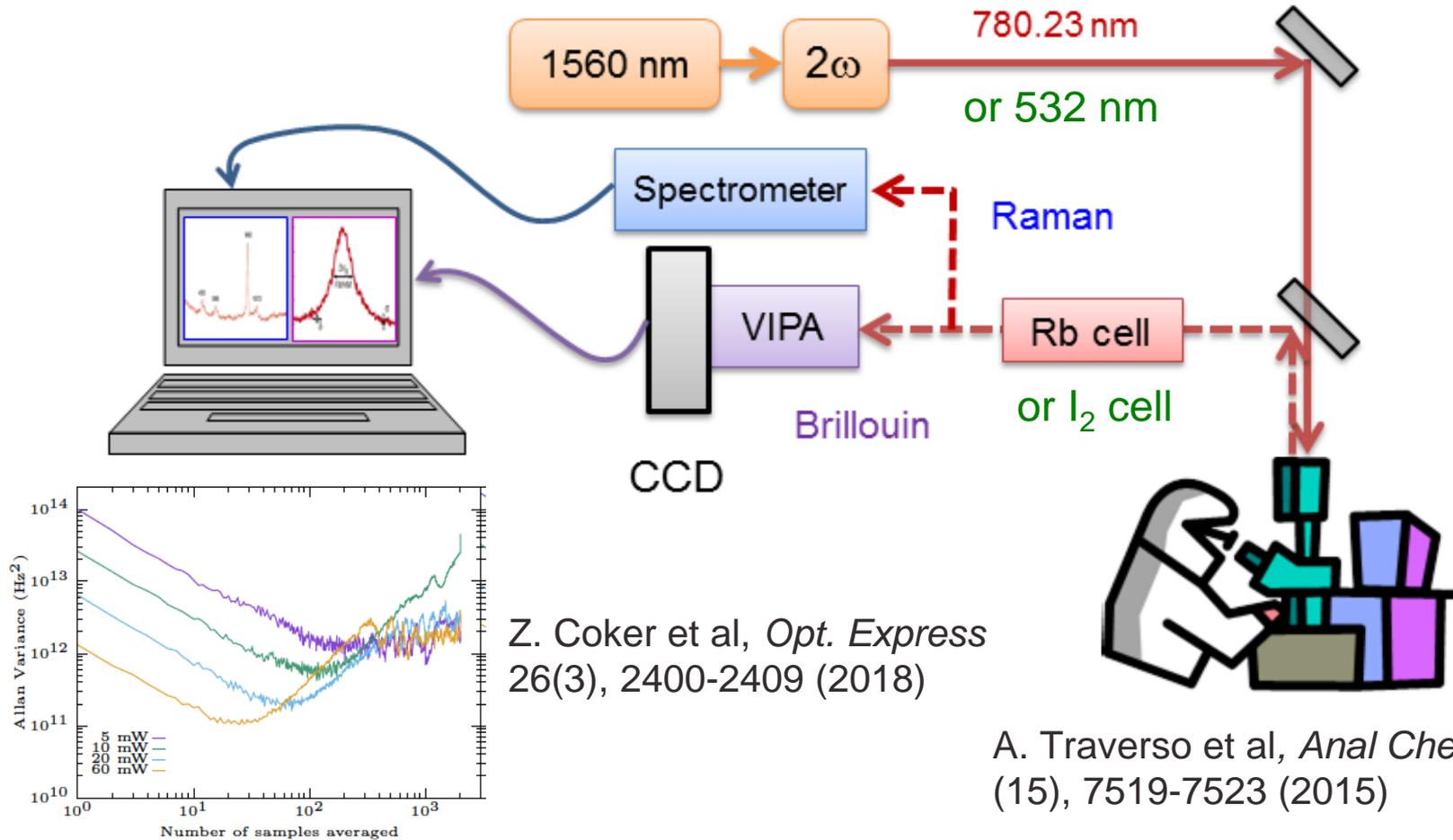
$$G^* = G' + i G'' = \rho v^2 + i (2\rho v^3 \alpha)$$

**Velocity** which is proportional to the **frequency shift**

**Attenuation** which is proportional to the **line-width**

Viscoelastic modulus is the function of frequency of acoustic wave, and, in Brillouin spectroscopy, we are assessing GHz waves. For nsPEF, those are the important ones.

# Mechanical assessment

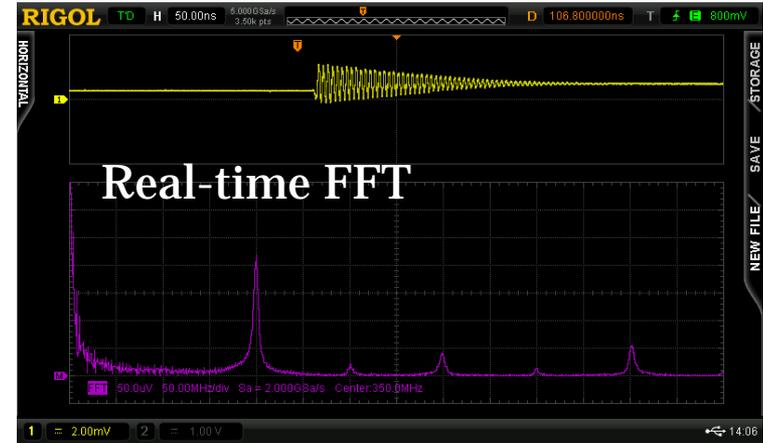
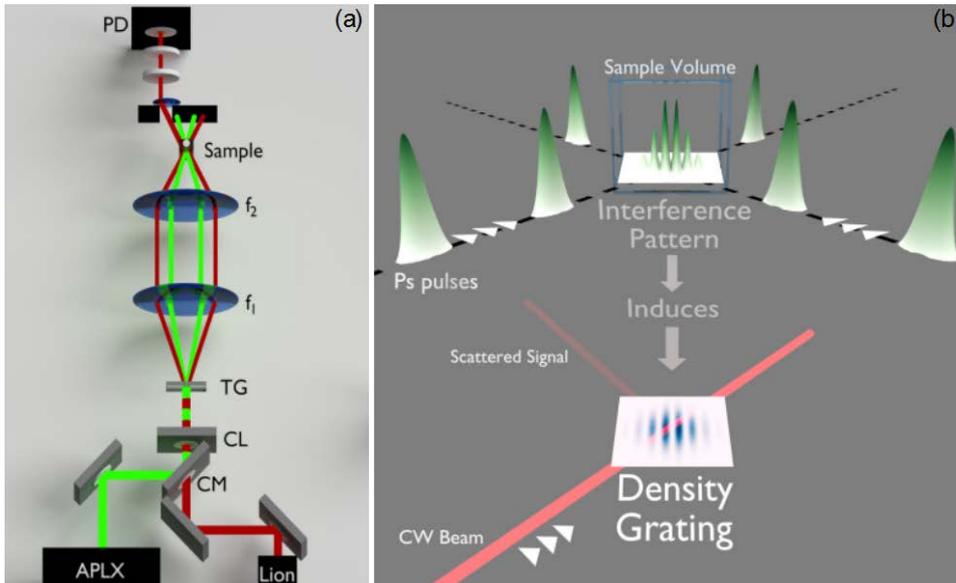


Z. Coker et al, *Opt. Express* 26(3), 2400-2409 (2018)

A. Traverso et al, *Anal Chem* 87 (15), 7519-7523 (2015)

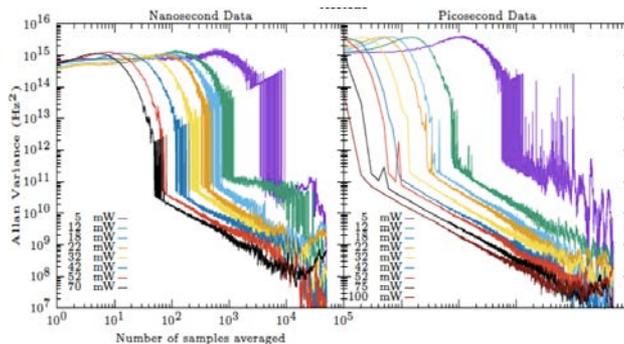
Zh. Meng, et al, *Adv. Opt. Phot.* 8(2), 300- 327 (2016).

# Mechanical assessment: BISTRO measurements



**B**rillouin **I**maging/**S**ensing via  
**T**ime-**R**esolved **O**ptical  
**(BISTRO)** Measurements

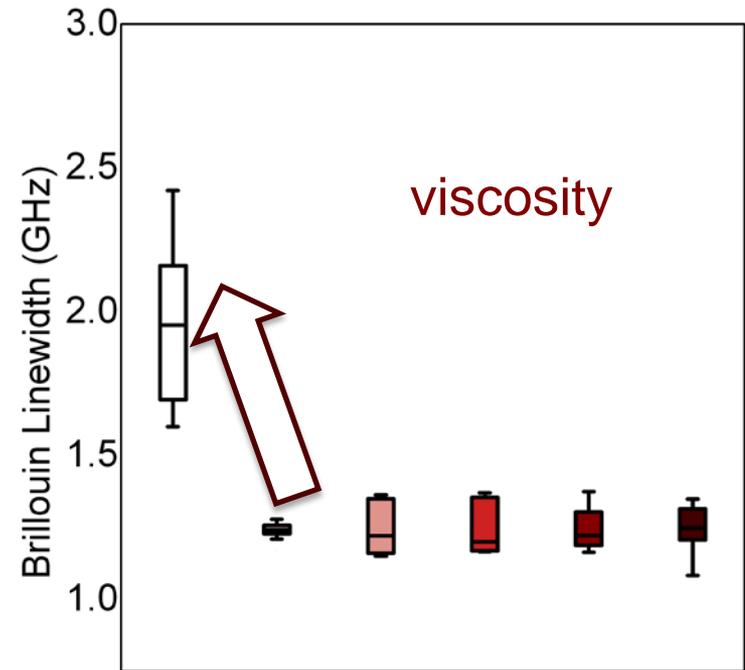
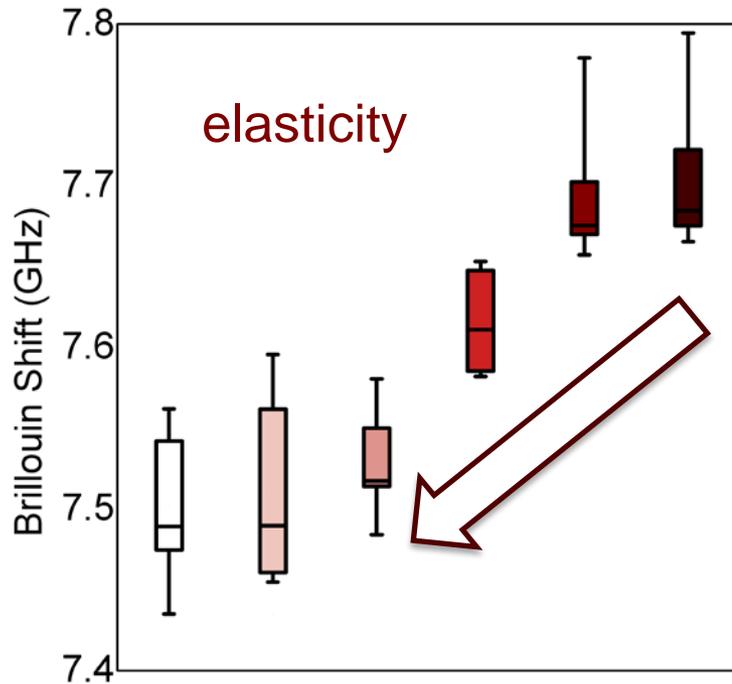
CW Ballmann, et al *Optica* 4(1), 124-128 (2017)



Shorter acquisition times (down to  $1\mu\text{s}$ )  
 More accurate measurements



# Mechanical assessment of nsPEF effects

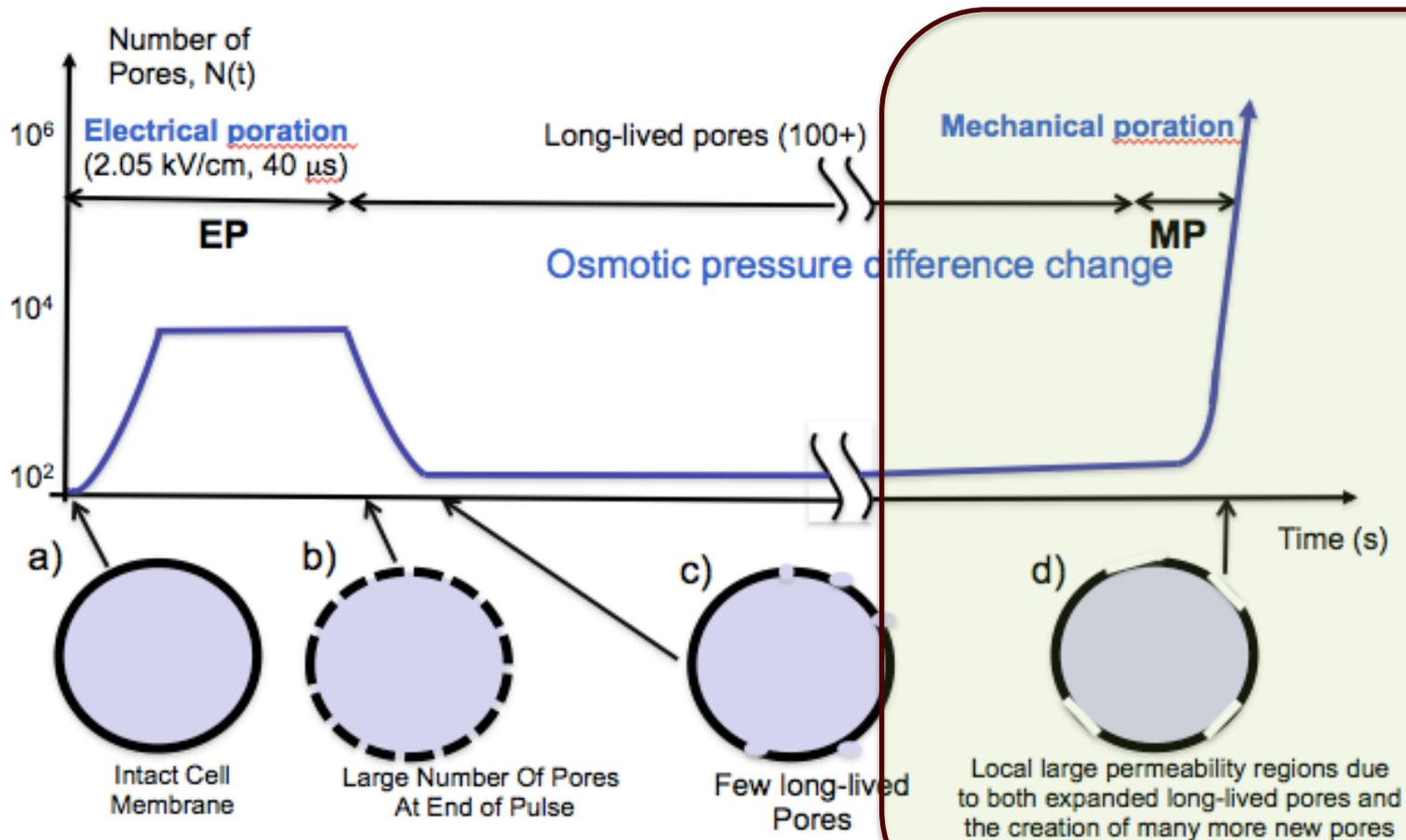


Number of nsEF pulses

Number of nsEF pulses

CHO (Chinese hamster ovarian) cells; 1.5 kV/cm, 600 ns pulse

# Comparison with theory



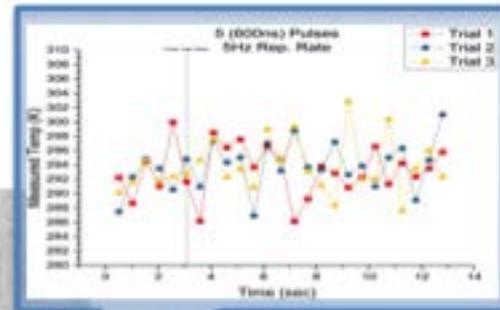
# Summary

Multiscale multimodal extension

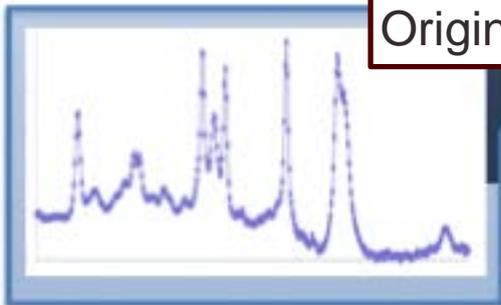
Electric field imaging



Temperature sensing

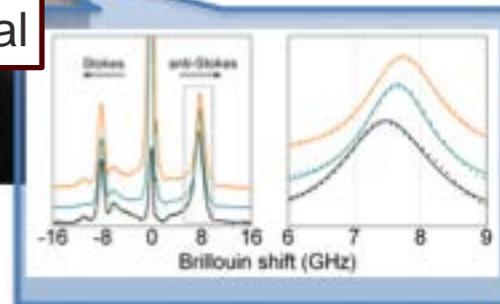


Chemical assessment



Original proposal

Mechanical assessment



# Sensitive Optical Fields' Imaging

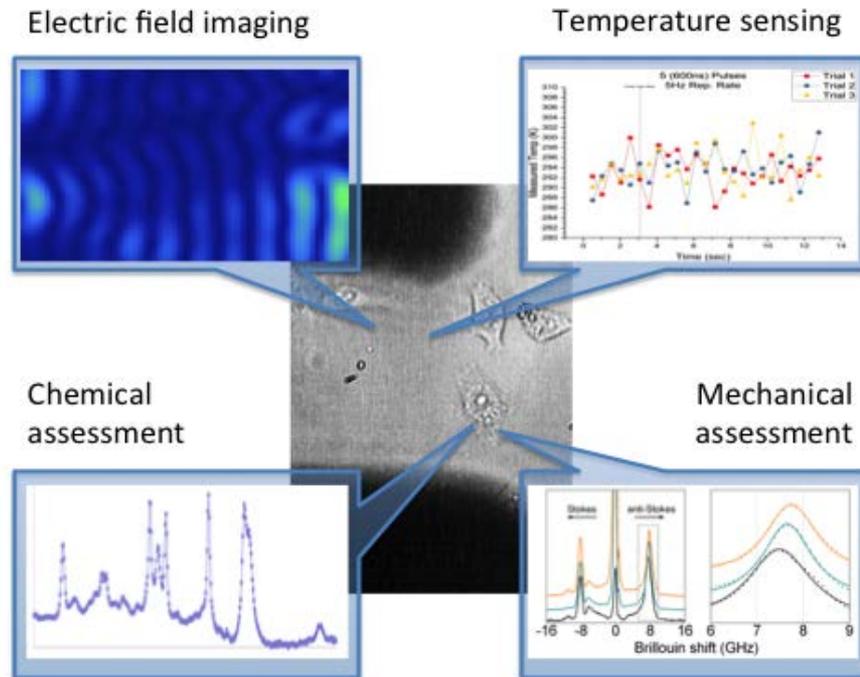
**Pockels effect:** possible at short time scales  
Provides with EF distribution  
Tomographic imaging makes  $E(x,y,z)$  possible

**CARS:** first experimental demonstration of Stark effect in model system of GUVs

**CARS:** system is optimized for high SNR measurements, so it is possible to start looking for small variations due to oxidation, electroporation, etc.

**NP:** "smart" substrate allows real-time temperature and magnetic field (due to electrical currents) sensing

**Diamonds NV:** imaging electrical currents and magnetic field (with Phil Hemmer)



**Brillouin:** sensitivity and signal strength are sufficient to look for dynamic changes immediately after nsPEF

# Future work

Year	1				2				3				4				5			
Quarter	1	2	3	4	1	2	3	4	1	2	3	4	1	2	3	4	1	2	3	4
<b>Task 4: Novel bio-electro-mechanical tools</b>																				
Develop broadband Raman imaging system																				
Develop coherent Brillouin imaging system																				
nsPEF experiments																				

**Nascent set of tools has been developed:** non-invasive microscopic assessment of electro-mechanical interactions in biological systems is now possible. State-of-the-art of sensitivity and accuracy measurements.

**Fast kinetics imaging:** pressing needs of nsPEF effects assessment.

- ✓ How fast is membrane discharge?
- ✓ What are the structure/chemistry changes associated with nsPEF?
- ✓ Are those changes permanent or transient?
- ✓ Are mechanical effects important?
- ✓ Theory vs Experiment. Can we connect those on molecular level?
- ✓ Any new physics?