



**AFOSR-FA9550-15-1-0326**

Experimental and theoretical investigation of the  
**Mechanisms of free-electron-mediated  
modification of biomolecules  
in nonlinear microscopy**

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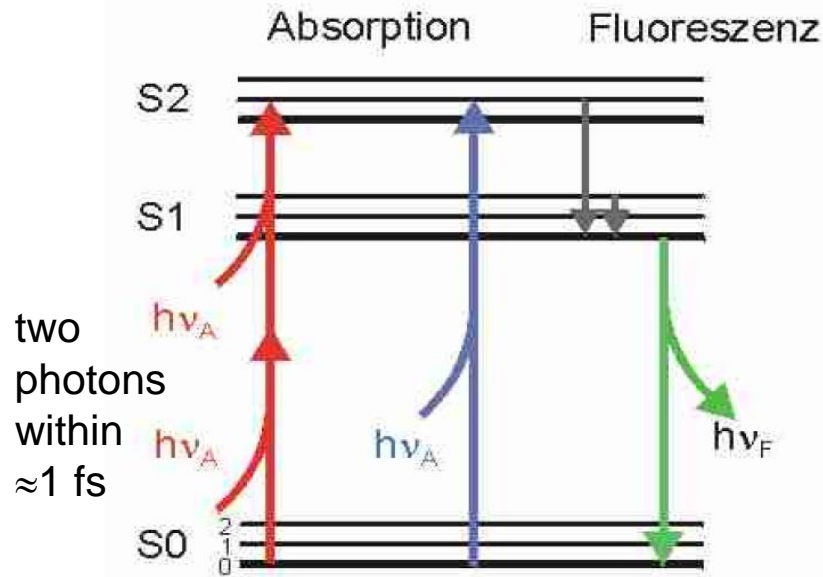
AFOSR Biophysics Program Review 2018-04-20

Public Release

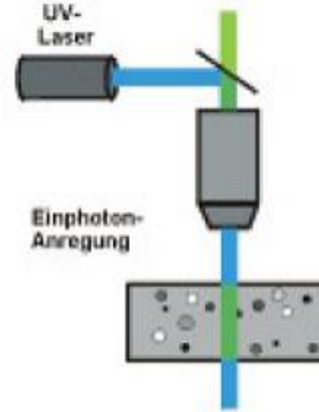
# Promise of multiphoton microscopy: Autofluorescence in-vivo imaging inside the body



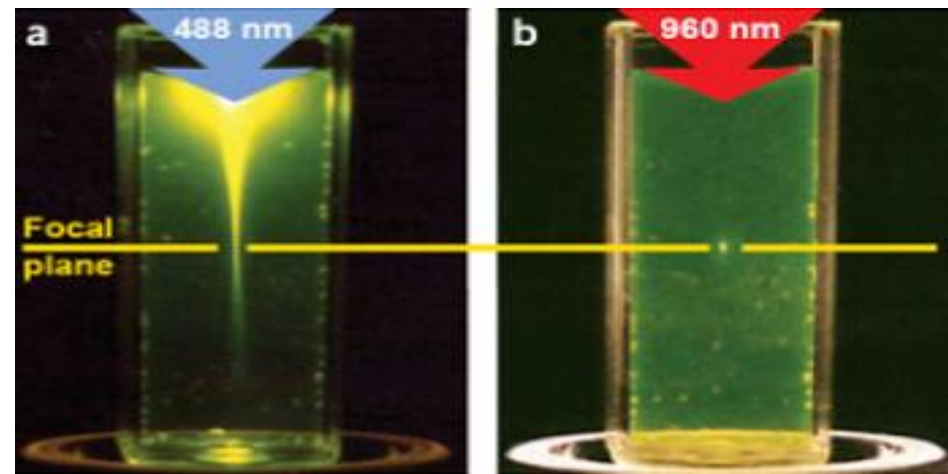
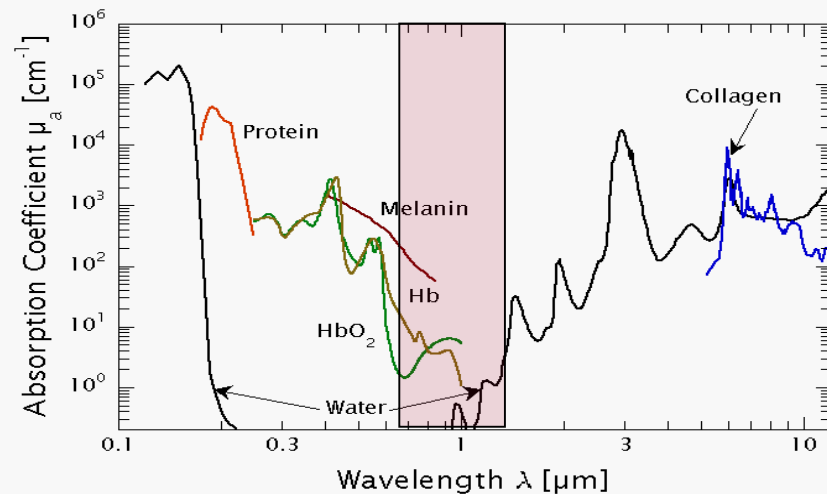
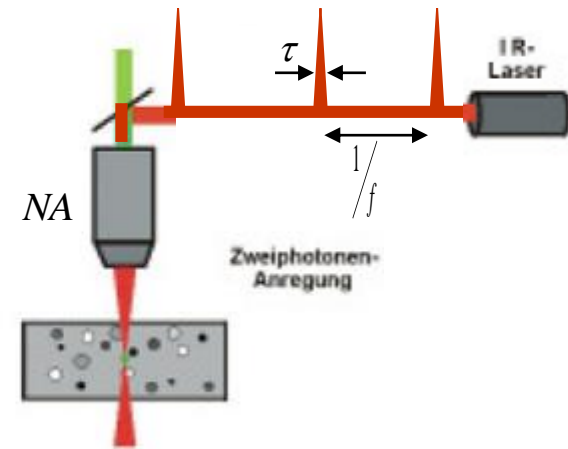
# Multiphoton microscopy



Cw UV laser  
(confocal microscopy)

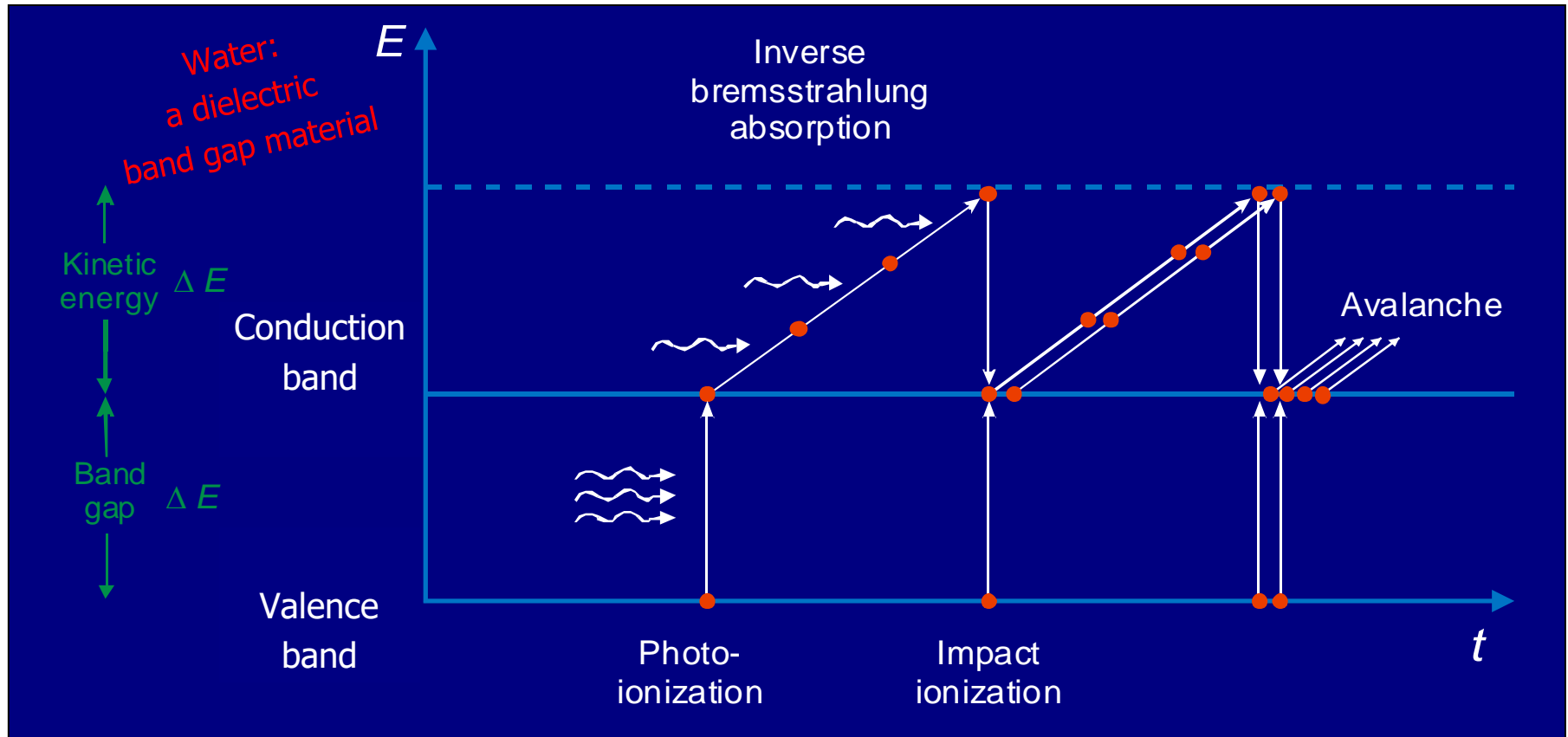


Pulse train (80 MHz) from femtosecond oscillator



- Up to 0.5 mm penetration depth into scattering tissue
- Well suited for in-vivo investigation of interfaces (skin, lung, intestine)

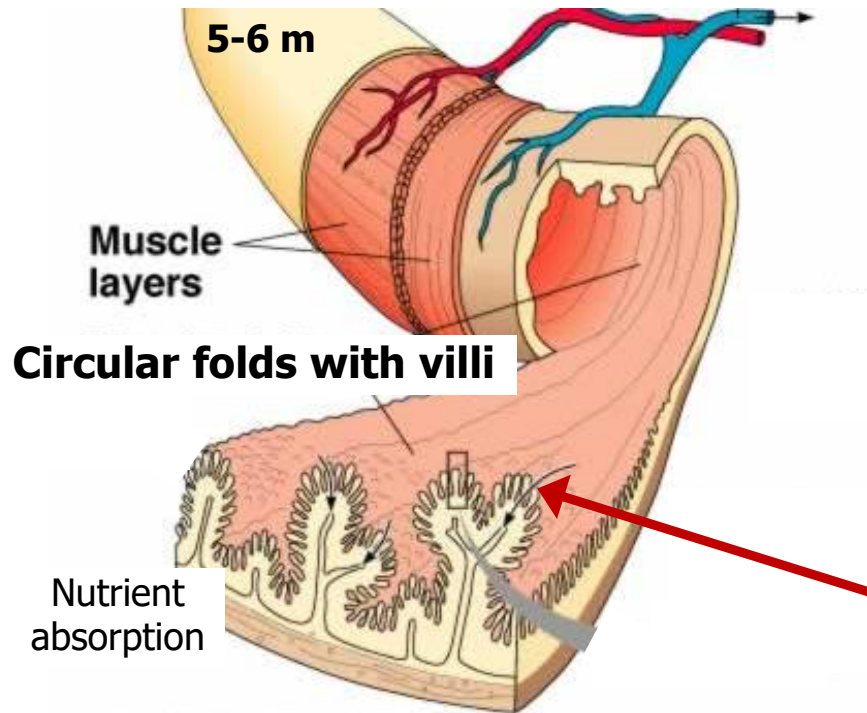
# Multiphoton excitation is not far away from plasma formation



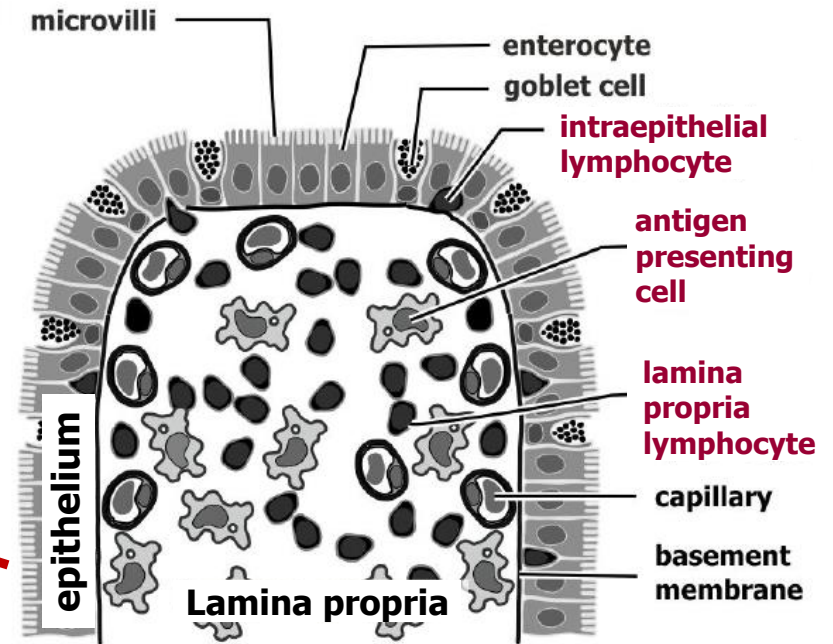
- High irradiance is needed for multiphoton excitation
- This implies a certain probability for multiphoton ionization
- That probability increases rapidly  $\propto I^k$  ( $k$  = order of multiphoton process)

# Example: Intestinal mucosa

## Small intestine



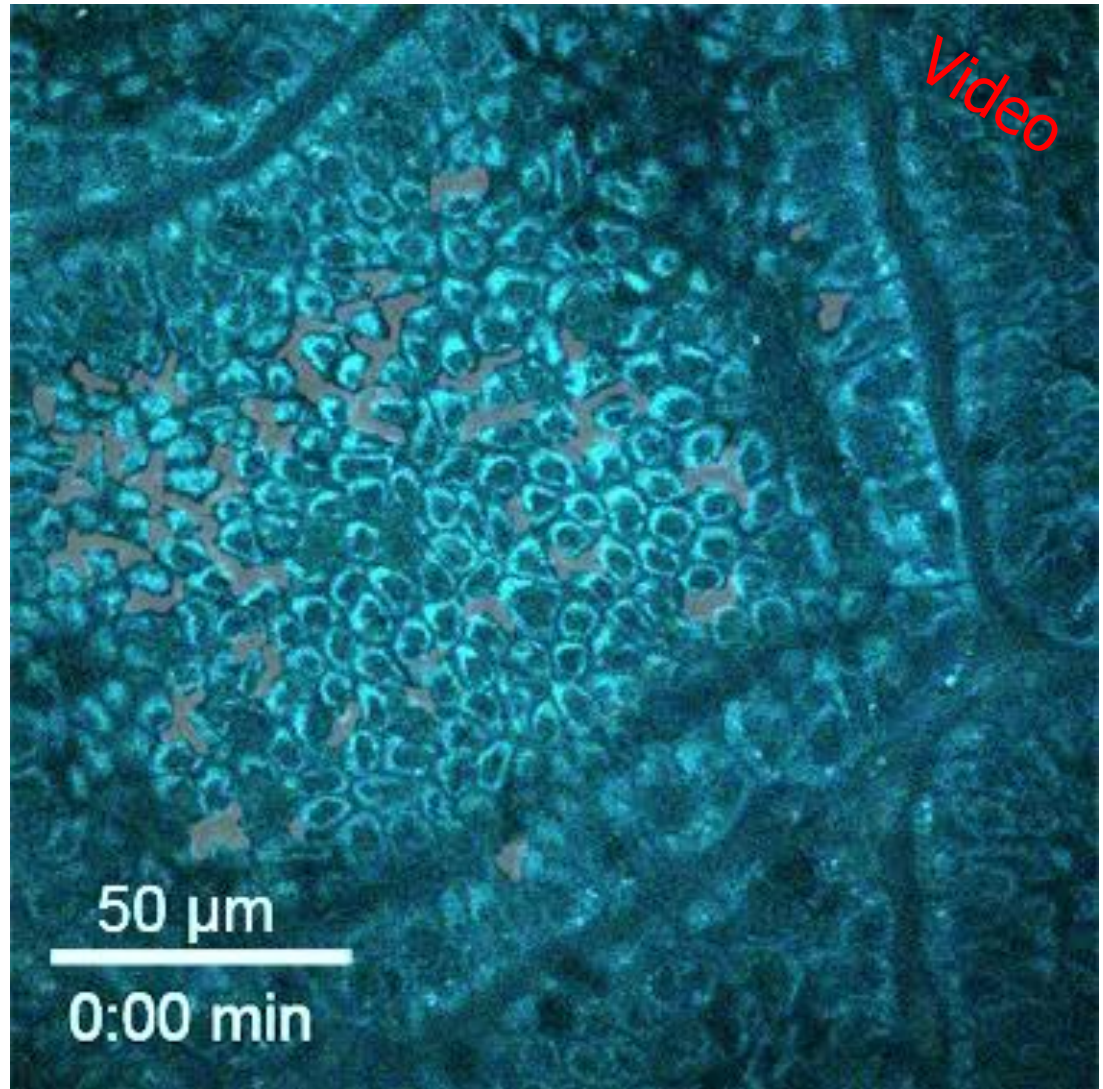
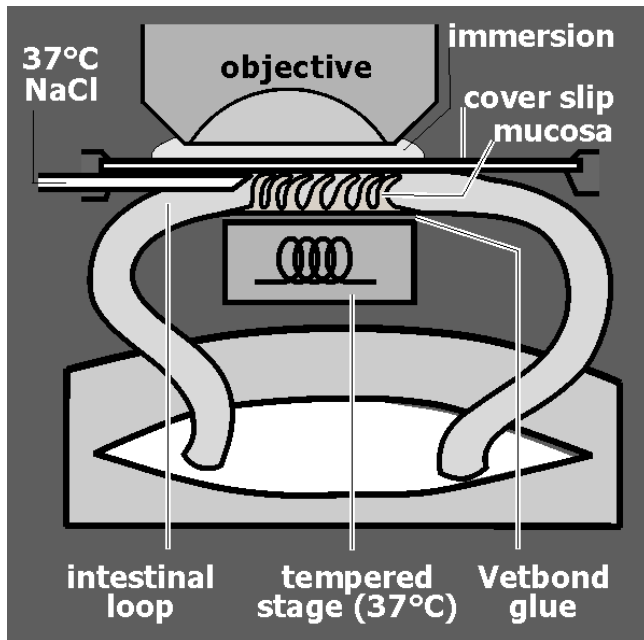
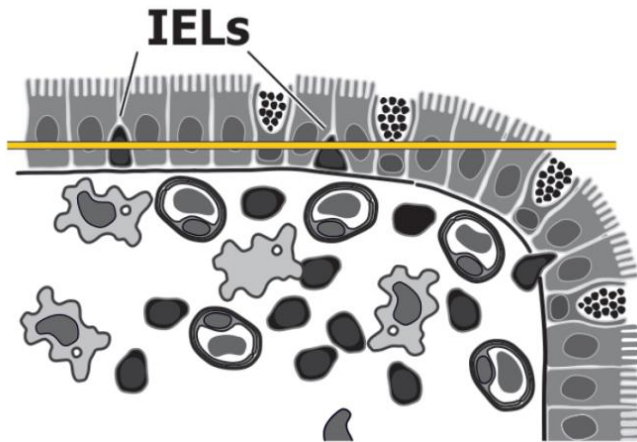
## Villus tip



Large surface ( $400\text{-}500\text{ m}^2$ )  
due to hierarchy of folds

Lymphocytes are parts of the  
adaptive immune system

# Movement of intraepithelial lymphocytes (within 13 min)

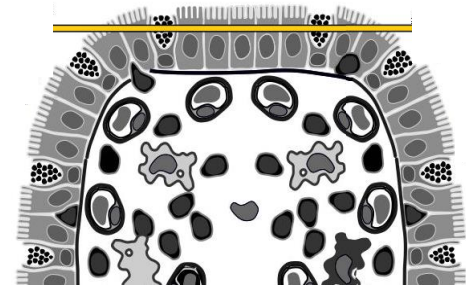
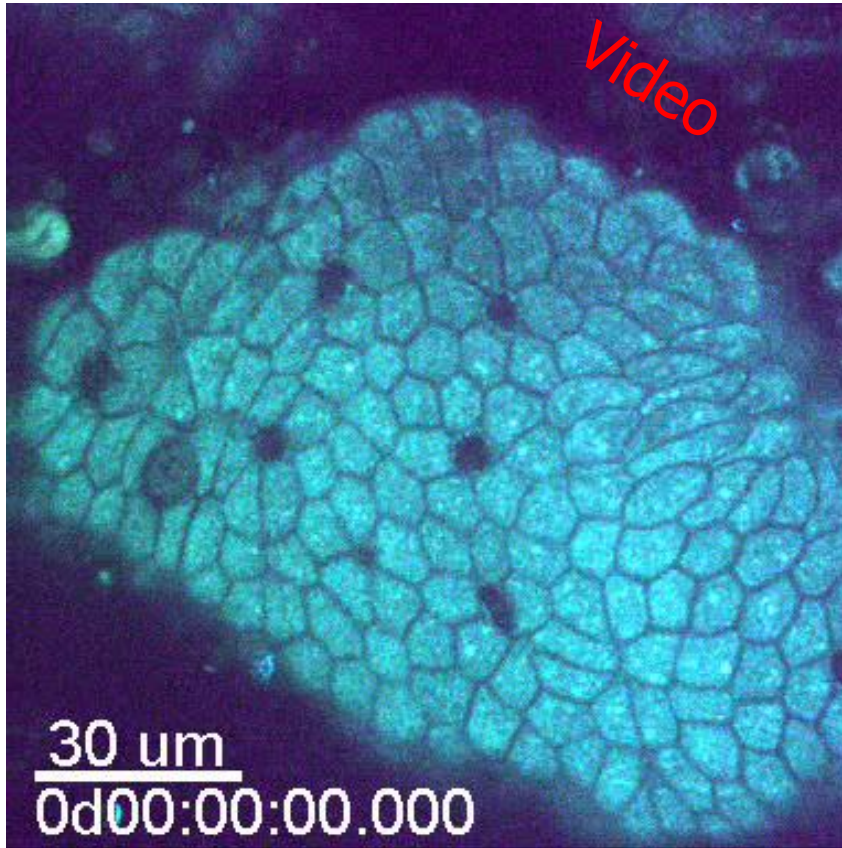


Orzekowsky-Schroeder et al. (2014)  
Biomed. Opt. Expr. 10: 3521-3540

The body's immune patrol in action

# Twice as much light → photodamage

Repeated (7x) scanning at 38 mW, 730 nm, NA = 1.2 (in total 7500 pulses/pixel)  
(2 x the power used for autofluorescence imaging)



## Hyperfluorescence

sets in at laser powers  
1.5-1.6 times larger  
than used for imaging

## Bubble formation

follows (dark spots in  
hyperfluorescent regions)

- Hyperfluorescence increases rapidly once started
- Onset of hyperfluorescence goes along with villus retraction

# Outline

**Imaging**

**Femtosecond pulse series**

- Evolution of „hyperfluorescence“ in different tissue types
- Is it hyperfluorescence – or plasma luminescence?
- Quantification of molecular disintegration via tracking of gas bubble formation



**Manipulation**

**Single-pulse effects**

# Nonlinear photochemistry @700-800 nm may introduce fluorescence changes

2-3 photon absorption

**Amino acids**

**NAD(P) H**

**Flavins**

**Porphyrins**



Hydroxyl  
radicals

$\text{H}_2\text{O}_2$

Reactive oxygen  
species (ROS)



Photoproducts with new fluorescence and  
nonlinear absorption properties

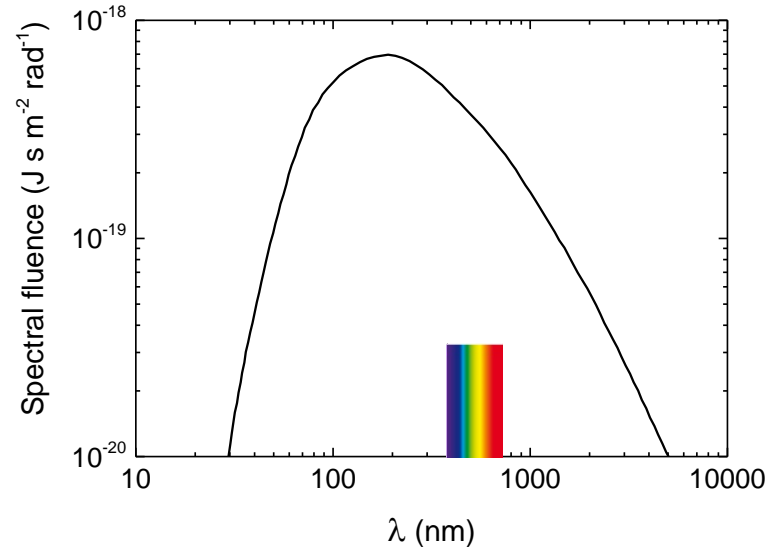
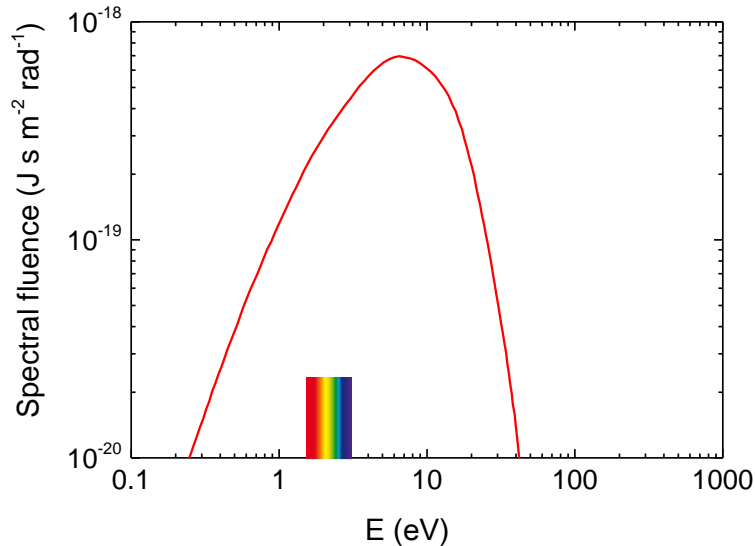


Functional damage, apoptosis, or necrosis

Laser-induced molecular changes may result in changes of  
fluorescence intensity and life time (detectable via FLIM).

# Free-electron formation $\Rightarrow$ plasma luminescence

Bremsstrahlung from the interaction of short laser pulses with dielectrics ( $\text{SiO}_2$ ) near breakdown threshold

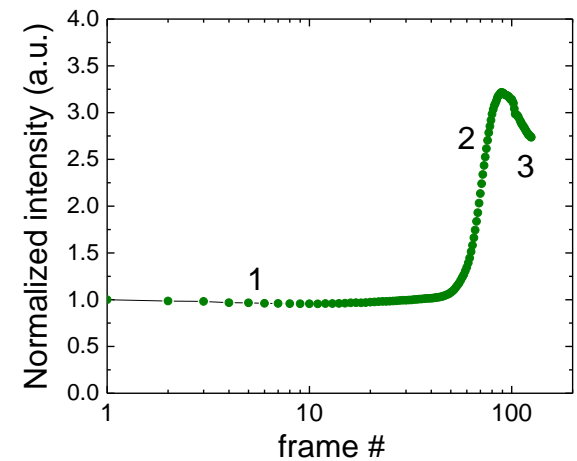
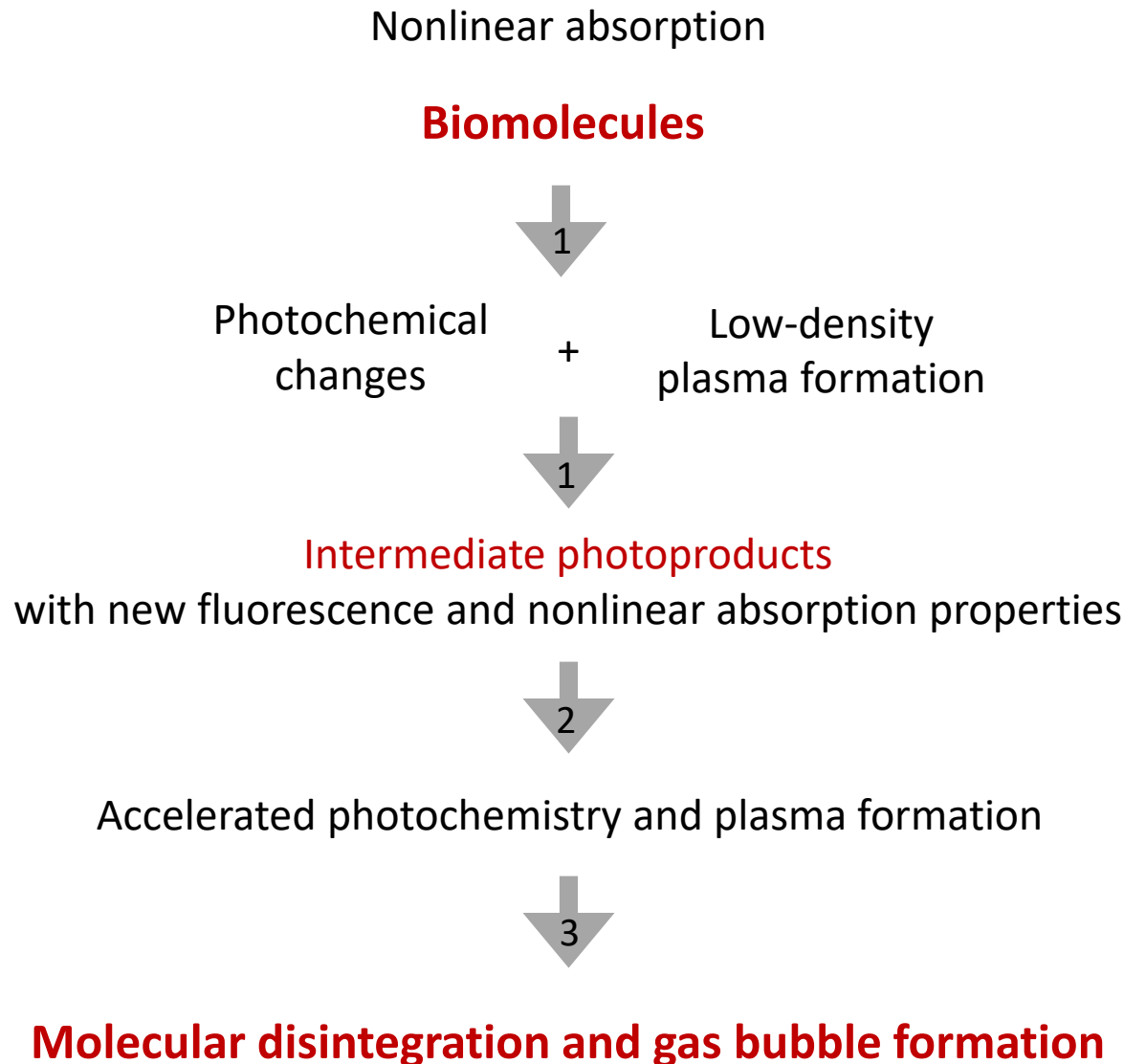


Adapted from

Petrov, Palastro & Peñano (2017) Bremsstrahlung from the interaction of short laser pulses with dielectrics, Phys. Rev. E 95, 053209, 1-10

- **Conduction band (CB) electrons with energies of a few eV emit visible Bremsstrahlung.** It becomes detectable when a burst of 80-MHz fs pulses is applied.
- **Recombination radiation** would be in the UV range ( $E_{\text{gap}} = 9.5 \text{ eV}$ ), and in water recombination is, moreover, largely non-radiative.
- **Blackbody radiation** is emitted from hot, thermalized plasma. However, the  $T$ -rise during multiphoton imaging is negligible. Bremsstrahlung is emitted before electron energies are thermalized.

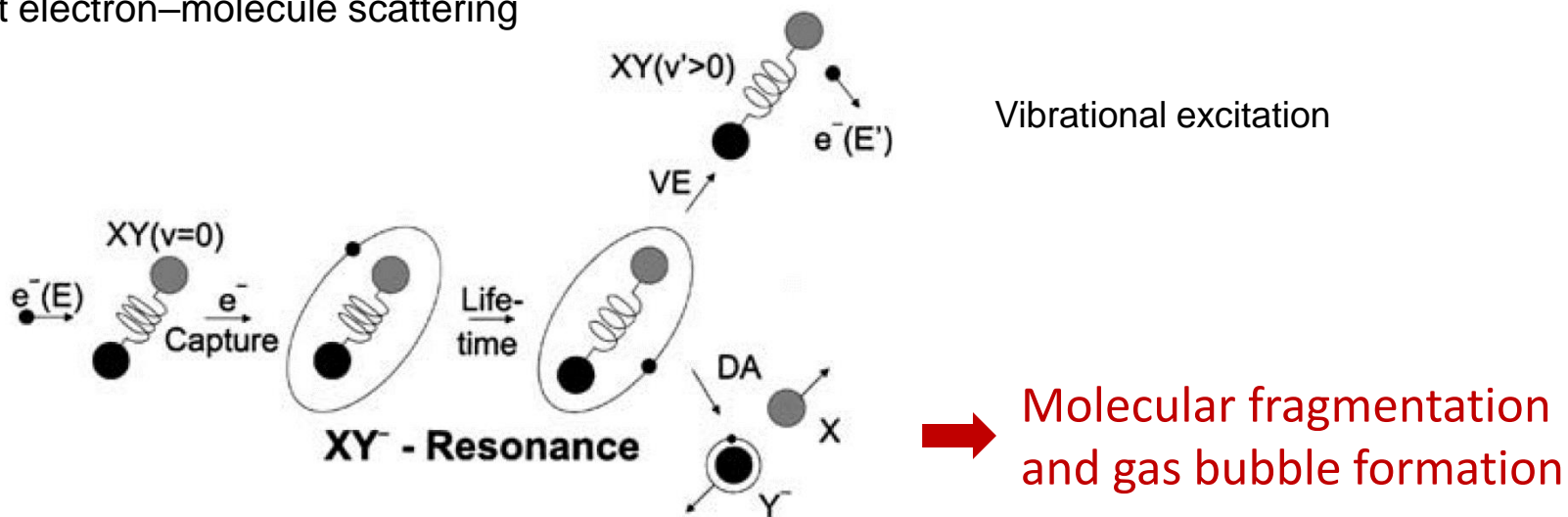
# Photodamage pathway suggested by the time evolution of fluorescence/luminescence and by FLIM



# Kinetics of gas bubble formation by single spot irradiation with fs laser pulse series

# Free-electron-induced molecular disintegration

**Dissociative electron attachment**  
in resonant electron–molecule scattering



Boudaiffa et.al. (2000) Science, 287:1658-1660

Vogel et al. (2005) Appl. Phys. B 81:1015–1047

- Free-electron mediated bond breaking → molecular fragmentation
- Cumulative molecular fragmentation during fs pulse series produces long-lived bubbles containing non-condensable gas (different from vapor bubble produced by a single laser pulse)

# Vapor bubbles in water $\Leftrightarrow$ gas bubbles in tissue

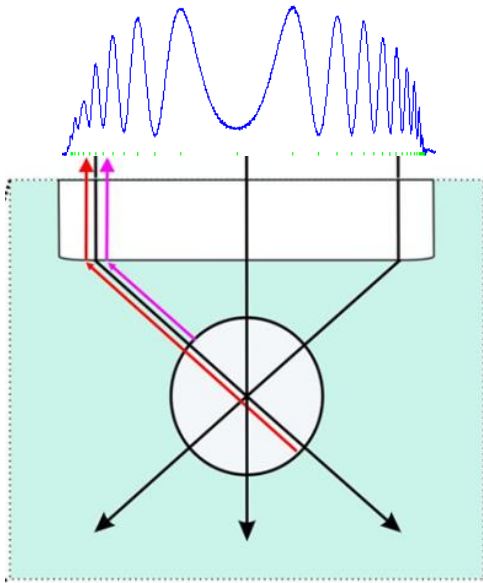
## Vapor bubble formation in water

- A bubble is formed, when the focal temperature exceeds the superheat threshold (spinodal limit)  $\Rightarrow$  phase explosion.
- After the phase explosion, the bubble expands beyond equilibrium and initially oscillates.
- Oscillations are damped, and an equilibrium between plasma-mediated vaporization and condensation evolves.
- The bubble disappears immediately at the end of the pulse series

## Gas bubble formation in cells & tissue

- Gas bubbles in tissue are formed by *cumulative* molecular disintegration.
- The **gas bubble threshold in cells** is much lower than the vapor bubble threshold in water. It **is closely linked to biomolecular changes**.
- The gas bubble grows continuously during the pulse series and vanishes via dissolution of the gas content  $\Rightarrow$  long bubble life time.
- **Quantification of gas formation bears info on molecular disintegration rates.**

# Measuring temporal evolution of bubbles from pulse series by combined interferometry and high-speed photography



- Bubble size evolution  $R(t)$  provides information on the rates at which biomolecules disintegrate into volatile products
- ⇒ Investigate  $R(t)$  for pulse series at various pulse energies as a function of wavelength



High-speed video  
at 100,000 frames/s

- High-speed photography provides a grid of reliable benchmark values for  $R$
- Interferometry provides precise info on the radius change  $dR/dt$

Radius evolution

$$R(t) = R_{Photo} + \int \dot{R}_{Interferometry} dt$$

One benchmark photo  
every 800 laser pulses  
in a 80 MHz train

# Evaluation of the mass of altered material in the bubble from the bubble volume

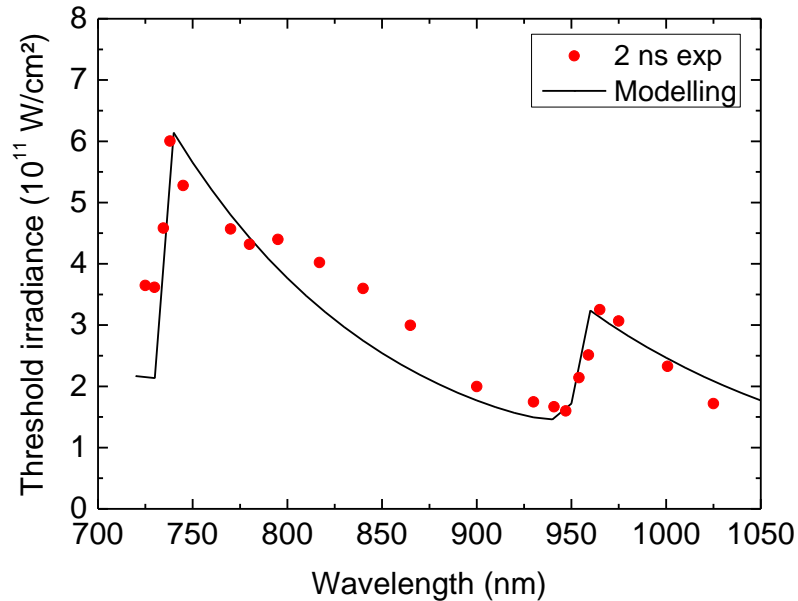
- Equilibrium between internal gas pressure and external pressure is assumed (This simplification is valid for small growth rates).
  - **Internal pressure** = gas pressure from molecular disintegration.
  - **External pressure** = hydrostatic pressure +  $p$ (surface tension) +  $p$ (restoring force of cellular matrix).
  - Mechanical properties of biological medium must be considered.
- 
- **Interferometry and modeling tools for single-shot-produced micro- and nano-bubbles ( $R_{\max} \rightarrow 0$ ) have already been developed (talk at last review meeting).**

Vapor bubble formation  
by single-pulse irradiation:

$$I_{\text{th}}(\lambda)$$

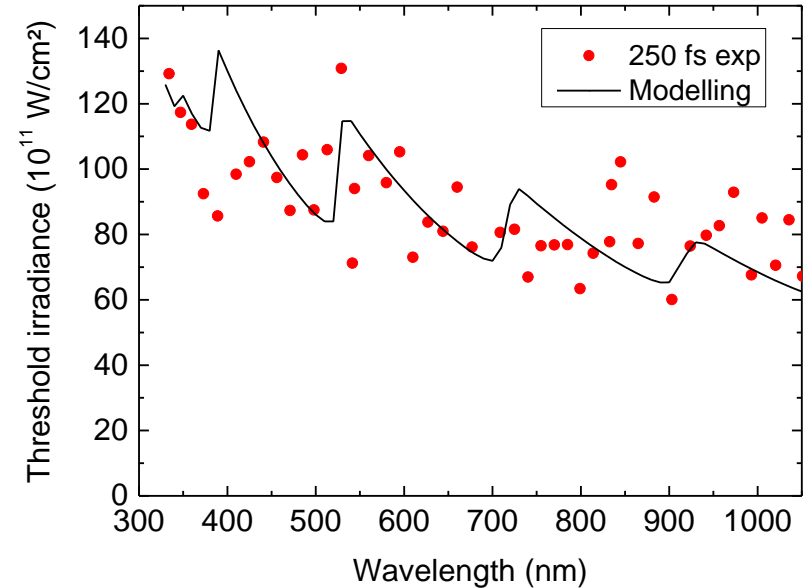
# Optical breakdown threshold spectra for single pulses in water

## Nanosecond breakdown



- Steps indicate multi-photon initiation
- Separation of the peaks proves the existence of an intermediate energy state  $E_{\text{ini}}$  at the solvated electron level that can be directly addressed from the valence band

## Femtosecond breakdown



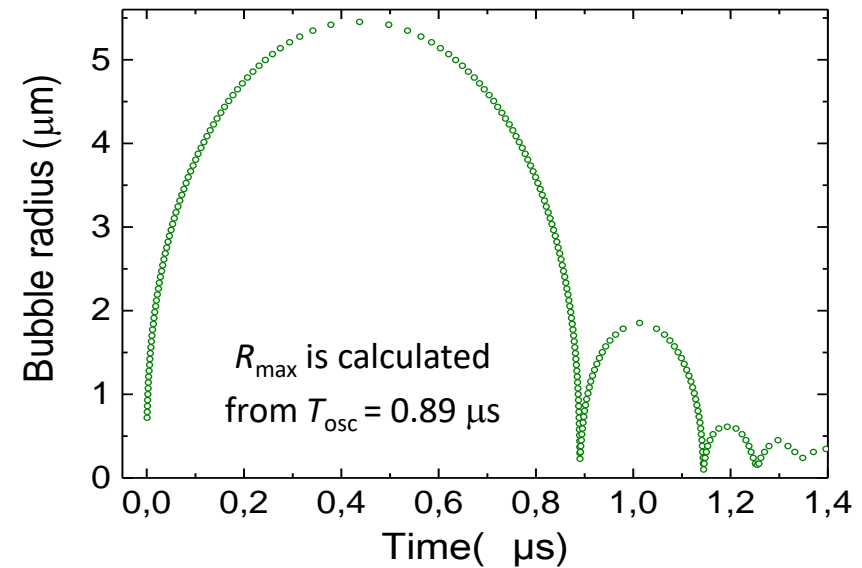
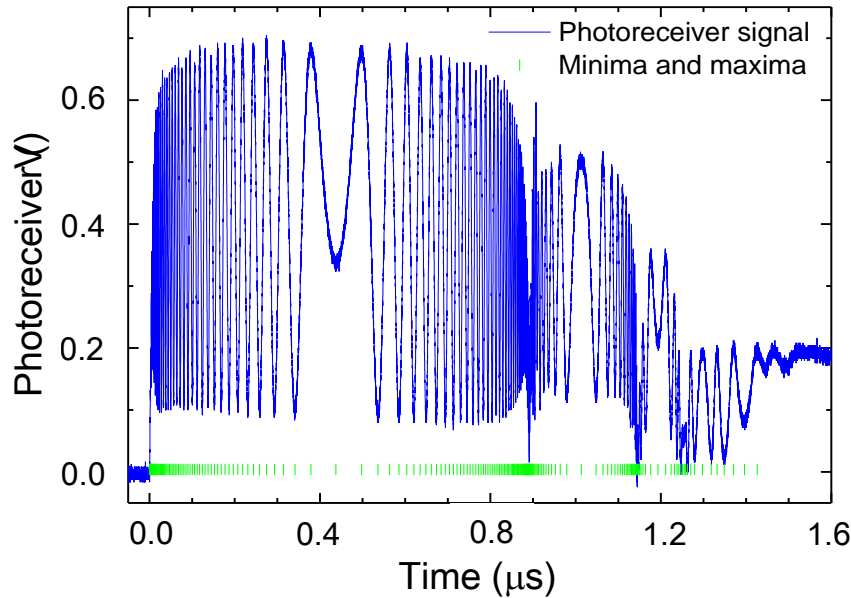
- Decrease of  $I_{th}$  with increasing  $\lambda$  indicates dominant role of avalanche ionization
- Breakdown model provides good fit for 1 fs effective Drude collision time

# Interferometry and modeling for single-shot produced bubbles

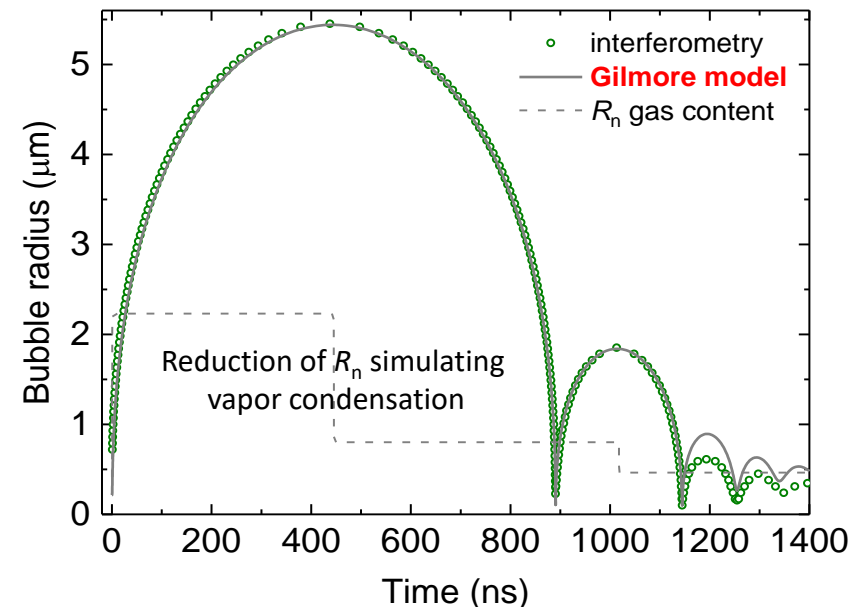
Interferometry signal

$R_{\max} = 5.4 \mu\text{m}$

$R(t)$  curve

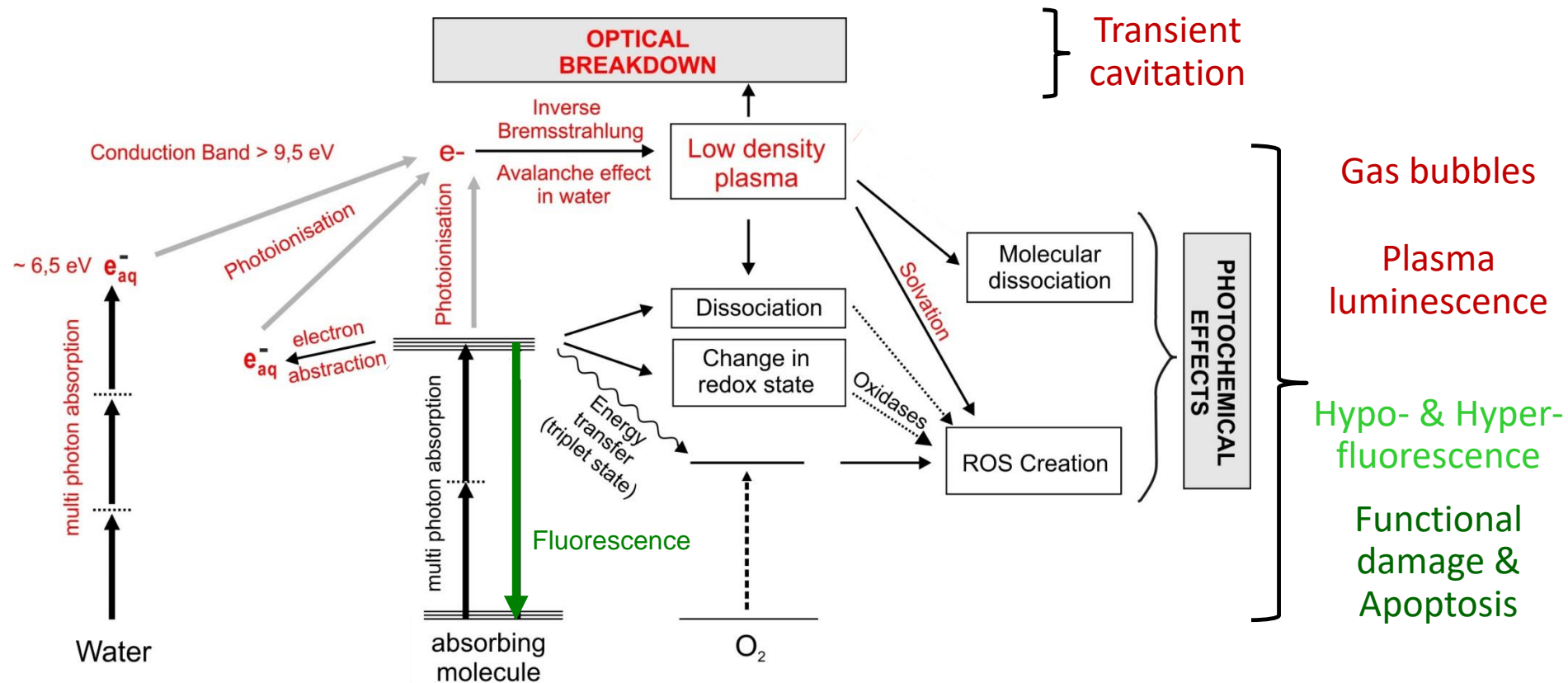


- Spatial resolution:  $< 5 \text{ nm}$   
Fringe max to min:  $70 \text{ nm}$
- Temporal resolution:  $160 \text{ ps}$
- Maximum detectable bubble wall velocity:  $440 \text{ m/s}$

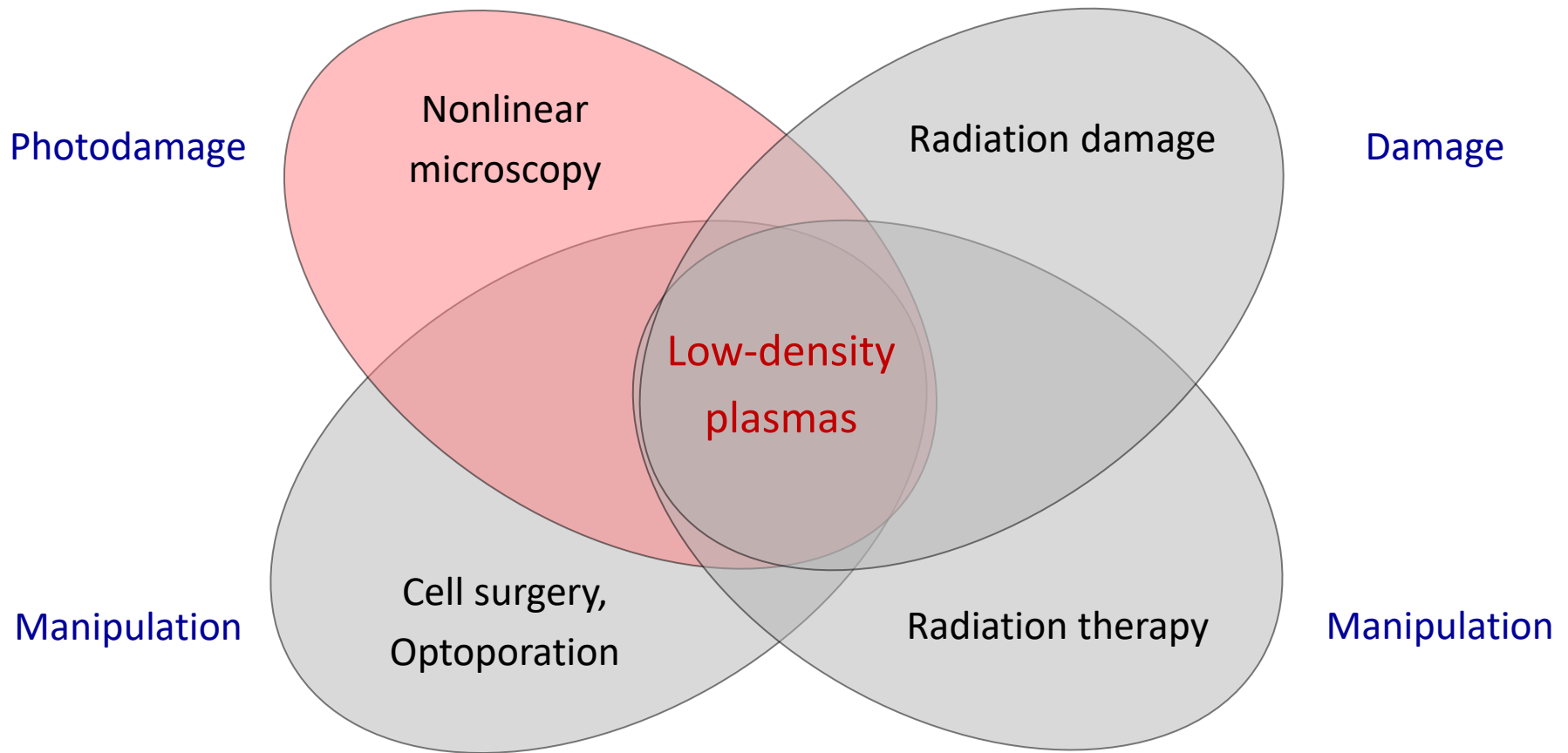


# Conclusions

# Pathways of nonlinear photo-modification of biomolecules in aqueous media



# Free-electron-mediated modifications



Free electrons are relevant as part of various **damage** mechanisms and can also be employed for **useful modifications**

# Conclusion

- We have collected more pieces of the puzzle describing modifications of biomolecules by nonlinear photochemistry and low-density plasma produced by MHz fs pulse series.



- A coherent picture of molecular interaction mechanisms will support optimum use of the photon budget in nonlinear microscopy, and establish new opportunities for the manipulation of biomolecules.

# Thank you from the project team

**PI: Alfred Vogel, FOSA, FSPIE**



**Co-PI: Norbert Linz**



**Experiments: Sebastian Freidank**



**Theoretical Modeling: Joe Liang**



**Institute of Biomedical Optics, University of Luebeck, Germany**